



# 3‑Fluoroazetidinecarboxylic Acids and trans,trans-3,4-Difluoroproline as Peptide Scaffolds: Inhibition of Pancreatic Cancer Cell Growth by a Fluoroazetidine Iminosugar

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# **S** [Supporting Information](#page-13-0)

ABSTRACT: Reverse aldol opening renders amides of 3-hydroxyazetidinecarboxylic acids (3-OH-Aze) unstable above pH 8. Aze, found in sugar beet, is mis-incorporated for proline in peptides in humans and is associated with multiple sclerosis and teratogenesis. Aze-containing peptides may be oxygenated by prolyl hydroxylases resulting in potential damage of the protein by a reverse aldol of the hydroxyazetidine; this, rather than changes in conformation, may account for the deleterious effects of Aze. This paper describes the synthesis of 3-fluoro-Aze amino acids as hydroxy-Aze analogues which are not susceptible



to aldol cleavage. 4-(Azidomethyl)-3-fluoro-Aze and 3,4-difluoroproline are new peptide building blocks. trans,trans-2,4-Dihydroxy-3-fluoroazetidine, an iminosugar, inhibits the growth of pancreatic cancer cells to a similar degree as gemcitabine.

# ■ INTRODUCTION

Organofluorine compounds play a central role in the pharma-ceutical industry.<sup>[1](#page-13-0)</sup> There is much current interest in azetidine,<sup>[2](#page-13-0)</sup> including azetidine carboxylic acids, $3$  as a pharmacophore in medicinal chemistry;<sup>[4](#page-13-0)</sup> over 180 patents with azetidine in the title have been published since 2013.<sup>[5](#page-13-0)</sup> Azetidinecarboxylic acid (Aze) 1, a four-membered ring analogue of proline 2 first isolated from *Convallaria majalis* in  $1955$ , <sup> $\delta$ </sup> occurs in many plants, including sugar beet,<sup>[7](#page-13-0)</sup> and in supplements to many foods.<sup>[8](#page-13-0)</sup> Aze is mis-incorporated into proteins as a substitute for proline 2 in humans and causes numerous toxic effects<sup>[9](#page-13-0)</sup> including teratogenesis and congenital malformations.[10](#page-13-0) There is a correlation between areas of high sugar beet consumption and the occurrence of multiple sclerosis; dangers of Aze entering the human food chain<sup>[11](#page-13-0)</sup> are increasingly recognized.<sup>[12](#page-13-0)</sup>

3-OH-Aze 3 has only been prepared by oxidation of Aze 1 by proline hydroxylases.<sup>[13](#page-14-0)</sup> The formation of trans,trans-3hydroxy-4-hydroxymethyl-L-Aze 4 from D-glucose was the first chemical synthesis of an unprotected 3-OH-Aze 3.<sup>[14](#page-14-0)</sup> Compound 3 is a ring-contracted analogue of the dihydroxyproline 5.<sup>[15](#page-14-0)</sup> The roles of 2-oxoglutarate (2OG)-dependent prolylhydroxylases in eukaryotes include collagen stabilization, hypoxia sensing, and translational regulation and are of considerable interest from therapeutic perspectives; the RPS23 hydroxylases in S. cerevisiae (Tpa1p), Schizosaccharomyces pombe, and green algae have been shown to catalyze an unprecedented dihydroxylation modification giving peptides containing 5. [16](#page-14-0) Synthetic peptides containing 5 or its azetidine analogues as constituents may provide interesting probes in biological investigations (Figure [1\)](#page-1-0). However, although the amide 6 is a potent inhibitor of hexosaminidases, $^{17}$  $^{17}$  $^{17}$  6 is unstable above pH 8 due to a reverse aldol opening of the azetidine ring and thus is not suitable for the preparation of stable peptidomimetics. Microbial 2-oxoglutaratedependent dioxygenases also cause oxidation of Aze-containing peptides 7 in which oxygen-modified hydroxyl-Aze peptides 8 will be susceptible to aldol ring opening, thus severely damaging the protein structure.<sup>[18](#page-14-0)</sup> Modification of the secondary structure of proteins containing Aze instead of Pro may affect their biological function; however, the damage in protein structure inherent in the introduction of oxygen into C3 of an azetidine amide may be the cause of much of the pathology of the Aze-Pro substitution. Access to stable functionalized Aze analogues may

Received: March 6, 2015 Published: April 10, 2015

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Figure 1. Azetidine and pyrrolidine structures: synthetic targets.

allow investigation of peptides with interesting biological and structural features.

Since unprotected 3-OH-Aze derivatives cannot give stable peptides, 3-fluoro-Aze 9 where fluorine replaces the hydroxyl group may be an alternative peptidomimetic. In 1970, 9 was formed by the direct photofluorination of Aze.<sup>[19](#page-14-0)</sup> In spite of there being  $\overline{\text{const}}$  considerable interest in 3-fluoroazetidines,<sup>20</sup> there are no reports of the use of 9 or its derivatives as a template for the introduction of 3-fluoro-Aze components into peptides.

This paper describes the synthesis of protected building blocks for the incorporation of the enantiomeric 3-fluoro-Aze 10L and 10D into peptides; both the amide 11 and a dimer derived from 10L were stable, showing 3-fluoro-Aze to be suitable as a peptide mimetic. Building blocks for inclusion of trans, trans-difluoroproline 12 and the  $\delta$ -amino-Aze 13 were also prepared; oligomers of  $\delta$ -amino-oxetane<sup>[21](#page-14-0)</sup> and -THF<sup>[22](#page-14-0)</sup> analogues of 13 show a predisposition toward adopting a wide range of secondary structures. None of the fluoro iminosugars, including the fluoroazetidines 14 and 15, and the difluoropyrrolidine analogue 16 of the  $\alpha$ -glucosidase inhibitor DAB 17, $^{23}$  $^{23}$  $^{23}$  showed any inhibition of any glycosidase. The meso-fluoroazetidine diol 15 was compared to 5-fluorouracil 18 and gemcitabine 19 and demonstrated significant inhibition of pancreatic cancer cell growth; the anticancer activity of fluoroazetidine 15 provides a further example of biological activity of iminosugars which does not depend on glycosidase inhibition.

#### ■ RESULTS AND DISCUSSION

Synthesis. Treatment of 1,3-di-O-triflates derived from carbohydrates with benzylamine provides a strategy for the efficient synthesis of many azetidines.<sup>[24](#page-14-0)</sup> In particular, the cyclization of 2,4-triflates of pyranoses, such as 22 and 25, to form bicyclic azetidines in high yields is reliable provided all the substituents in the pyranose ring are equatorial (trans to each other).[25](#page-14-0) No other stereochemistry allows any cyclization, and it has not been possible to adjust the stereochemistry after the bicyclic azetidine has been formed.<sup>[26](#page-14-0)</sup>

The diacetonide of 3-fluoroglucose 21, the starting material for all the 3-fluoroazetidines prepared, is formed in 97% yield from diacetone allose  $20^{27}$  $20^{27}$  $20^{27}$  by triflation and nucleophilic displacement

with cesium fluoride in 2-methyl-2-propanol<sup>[28](#page-14-0)</sup> on a large scale (over 100 g). $^{29}$  $^{29}$  $^{29}$  Compound 21 was transformed by standard carbohydrate manipulations into the pyranose ditriflates 22 and 25 which underwent high-yield reactions with benzylamine to give the bicyclic azetidines 23 and 26, respectively. Ring opening of 23, followed by periodate cleavage of the C5−C6 bond of glucose, allowed access to the fluoro-D-Aze 24D. The bicyclic azetidine 26, in which the C5−C6 bond of glucose had been cleaved prior to cyclization, led to the L-enantiomer 24L. Activation of the primary hydroxyl group in 24L toward nucleophilic substitution can lead to the synthesis of the azidomethyl-Aze 27 or of the ring-expanded difluoroproline 28 as building blocks for the incorporation of novel amino acids into peptides (Scheme [1](#page-2-0)).

The synthesis of the D-trans,trans-Aze ester 24D from the diacetonide 21 required cleavage of the C5−C6 bond after the formation of azetidine ring (Scheme [2\)](#page-2-0). Hydrolysis of the acetonide  $21$  by Dowex  $H^+$ , followed by peracetylation in pyridine, afforded the tetraacetate 29 as a 1:1 anomeric mixture  $(97%)$ <sup>[30](#page-14-0)</sup> Treatment of the tetraacetate 29 with HBr in acetic acid gave the anomeric bromide which, with methanol in the presence of silver carbonate, gave the  $\beta$ -methyl pyranoside 30<sup>[31](#page-14-0)</sup> (87%). Removal of the acetate groups in 30 with sodium methoxide in methanol gave the triol 31 (96%) in which the primary hydroxyl group selectively reacted with tert-butyldimethylsilyl (TBDMS) chloride in the presence of imidazole in DMF to give the silyl ether 32 (98%). Esterification of the diol 32 with trifluoromethanesulfonic (triflic) anhydride formed the ditriflate 22. Treatment of 22 with benzylamine in the presence of N,Ndiisopropylethylamine (DIPEA) in acetonitrile afforded the bicyclic azetidine 23 (84% over two steps); only the  $\beta$ -anomer of the ditriflate 22 allowed cyclization to the azetidine ring.

The acid hydrolysis of the bicycle 23 under a wide range of conditions, followed by treatment of the crude product with sodium borohydride, only gave traces of the triol 34. However, acetolysis of 23 by acetic anhydride in the presence of boron trifluoride etherate gave the mixed acetal 33 in 100% yield. All attempts to directly reduce 33 with sodium borohydride under a variety of conditions afforded only traces of 34. Accordingly sequential reduction of the triacetate acetal 33 with DIBALH in toluene, followed by sodium borohydride in methanol, formed

<span id="page-2-0"></span>Scheme 1. Synthetic Strategy and Targets (Numbering Refers to Original C in Glucose)



Scheme 2. Synthesis of D-trans, trans-Aze  $24D^a$ 



<sup>a</sup>Reagents and conditions (i) Dowex (50W X8, H<sup>+</sup>), then Ac<sub>2</sub>O, pyridine, 97% (ii) HBr, AcOH, then AgCO<sub>3</sub>, MeOH, 87%; (iii) NaOMe, MeOH, 96%; (iv) TBDMSCl, imidazole, 98%; (v) (CF<sub>3</sub>SO<sub>2</sub>)<sub>2</sub>O, pyridine; (vi) BnNH<sub>2</sub>, DIPEA, MeCN, 84%; (vii) Et<sub>2</sub>O·BF<sub>3</sub>, Ac<sub>2</sub>O, 100%; (viii) DIBALH, toluene, then NaBH<sub>4</sub>, MeOH, 97%; (ix) Pd/C, H<sub>2</sub>, H<sub>2</sub>O, dioxane, 88%; (x) NaIO<sub>4</sub>, H<sub>2</sub>O, dioxane, then I<sub>2</sub>, K<sub>2</sub>CO<sub>3</sub>, MeOH, 69%; (xi) K<sub>2</sub>CO<sub>3</sub>, H<sub>2</sub>O, dioxane, 57%; (xii) Pd/C, H<sub>2</sub>, H<sub>2</sub>O, dioxane, 45% (numbering refers to original C in glucose).

the azetidine iminosugar 34 in 97% yield. These conditions were also required in the hydrolysis of other bicyclic nitrogen heterocycles.<sup>[32](#page-14-0)</sup> Oxidative cleavage of the 1,2-diol moiety in 34 by sodium periodate in aqueous dioxane, followed by further oxidation of the crude aldehyde by iodine in methanol, gave the peptidomimetic building block 24D in 69% yield (45% overall yield from the diacetone fluoroglucose 21).

Unprotected azetidine derivatives were prepared from 24D for assaying their biological properties. Hydrogenolysis of the benzyl group in 34 in the presence of 10% palladium on charcoal in aqueous dioxane gave the azetidine iminosugars 14 (88%). Hydrolysis of the methyl ester 24D by potassium carbonate in aqueous dioxane gave the N-benzyl Aze  $35D(57%)$  from which the benzyl group was removed by hydrogenolysis in the presence of palladium to give the fluoro D-Aze 10D (45%).

A similar sequence of reaction with cleavage of the C5−C6 bond before the formation of azetidine ring gave the L-trans, trans-Aze

ester 24L (Scheme [3](#page-3-0)). Mild acid hydrolysis of the diacetonide 21 gave the monoacetonide 36 (100%). Oxidation of the diol in 36 by sodium periodate, followed by sodium borohydride reduction of the resulting aldehyde, afforded 37 (92%). Hydrolysis of 37 by Dowex resin in aqueous dioxane gave 3-fluoroxylose 38<sup>[33](#page-14-0)</sup> (88%) in an  $\alpha$ : $\beta$  ratio of 3:2. Compound 38 with acetic anhydride in pyridine yielded an anomeric mixture of the triacetate 39 (89%). Treatment of 39 with HBr in acetic acid followed by reaction of the resulting bromide with silver carbonate in methanol gave the diacetate 40 (60%); removal of the acetate protecting groups by sodium methoxide in methanol gave the β-methyl xylopyranoside 41 (97%). Esterification of the diol 41 with triflic anhydride in dichloromethane in the presence of pyridine gave the ditriflate 25; treatment of 25 with benzylamine in acetonitrile in the presence of DIPEA afforded the bicyclic azetidine 26 (97%). Hydrolysis of the acetal in 26 with hydrochloric acid in aqueous dioxane gave the corresponding lactol <span id="page-3-0"></span>Scheme 3. Synthesis of L-trans,trans-Aze  $24L^a$ 



a<br>Reagents and conditions (i) MeOH, 1% H<sub>2</sub>SO<sub>4</sub>, 100%; (ii) NaIO<sub>4</sub>, H<sub>2</sub>O, dioxane, then NaBH<sub>4</sub>, 92%; (iii) Dowex (50W X8, H<sup>+</sup>), H<sub>2</sub>O, dioxane, 88%; (iv) Ac<sub>2</sub>O, pyridine, 89%; (v) HBr, AcOH; then AgCO<sub>3</sub>, MeOH, 60%; (vi) MeONa, MeOH, 97%; (vii) (CF<sub>3</sub>SO<sub>2</sub>)<sub>2</sub>O, pyridine, CH<sub>2</sub>Cl<sub>2</sub>; then BnNH<sub>2</sub>, MeCN, 97%; (viii) HCl, H<sub>2</sub>O, dioxane; then  $I_2$ , K<sub>2</sub>CO<sub>3</sub>, MeOH, 74%; (ix) K<sub>2</sub>CO<sub>3</sub>, H<sub>2</sub>O, dioxane, 43%; (x) Pd/C, H<sub>2</sub>, H<sub>2</sub>O, dioxane, 82%; (xi) HCl, H<sub>2</sub>O, dioxane; then NaBH<sub>4</sub>, MeOH, 40%; (xii) NaBH<sub>4</sub>, MeOH, 90%; (xiii) Pd/C, H<sub>2</sub>, H<sub>2</sub>O, dioxane, 100% (numbering refers to original C in glucose).

#### Scheme 4. Peptide Building Blocks<sup>a</sup>



a<br>Reagents and conditions: (i) MsCl, pyridine, 100%; (ii) NaN3, DMF, 80%; (iii) XtalFluor-M, Et3N·3HF, CH2Cl2, 84%; (iv) NaOH, H2O, dioxane, 64%; (v) Pd/C, H2, H2O, dioxane, 86%; (vi) LiAlH4, THF, 66%; (vii) Pd/C, H2, H2O, dioxane, 82%.

which was oxidized by iodine in methanol to give the L-Aze ester building block 24L (74%) in an overall yield of 30% from the diacetonide 21.

Hydrolysis of the methyl ester 24L by potassium carbonate in aqueous dioxane gave the N-benzyl acid 35L (43%) from which the benzyl group was removed by hydrogenolysis in the presence of palladium on charcoal to produce the azetidine carboxylic acid

10L (82%). The lactol derived from hydrolysis of 26 was reduced by sodium borohydride in methanol to give the meso-diol 42 (40%); alternatively, sodium borohydride reduction of the ester 24L gave 42 in 90% yield. Hydrogenolysis of the N-benzyl group in 42 gave the meso-azetidine 15 (100%).

Other peptide building blocks were prepared from the L-ester 24L (Scheme 4). The mesylate 43, formed by treatment of 24L <span id="page-4-0"></span>Scheme 5. Amide Formation<sup>a</sup>



<sup>a</sup>Reagents and conditions: (i) MeNH<sub>2</sub>, CaCl<sub>2</sub>, MeOH, 74%; (ii) Pd/C, H<sub>2</sub>, H<sub>2</sub>O, dioxane, 90%; (iii) TBDMSCl, imidazole, DMF, 95%; (iv) MeNH<sub>2</sub>, CaCl<sub>2</sub>, MeOH, 100%; (v) K<sub>2</sub>CO<sub>3</sub>, H<sub>2</sub>O, dioxane; (vi) Pd/C, H<sub>2</sub>, H<sub>2</sub>O, dioxane; (vii) 51, 53, HBTU, DMF, 45% from 50.



Figure 2. Compounds assayed for inhibition of glycosidases and/or pancreatic cancer cell growth.

with mesyl chloride in pyridine (100%), with sodium azide in DMF gave the azide 27 (80%) as a precursor to  $\delta$ -amino Aze analogues. A variety of reagents for attempted displacement of the mesylate 43 by fluoride nucleophiles gave complex mixtures, which gave very low yields of the difluoroazetidine 46. However, reaction of 24L with XtalFluor-M<sup>[34](#page-14-0)</sup> and triethylamine:HF in dichloromethane afforded the difluoroproline ester 28 (84%). Neighboring group participation by the ring nitrogen on the initial complex 44 with the azetidine nitrogen would give the bicyclic aziridinium ion 45 to afford the ring-expanded proline 28. Compound 28 would allow fluoro analogues of dihydroxyproline 5 to be incorporated into peptides. Examples of ring expansions of azetidines to pyrrolidines<sup>[35](#page-14-0)</sup> and opening of aziridinium ions by fluoride $36$  have been reported.

Hydrolysis of 28 with aqueous sodium hydroxide gave the Nbenzylproline 48 (64%) from which the benzyl group was removed by palladium-catalyzed hydrogenolysis to give trans, trans-3,4-difluoro-L-proline 12 (86%). Reduction of 28 with lithium aluminum hydride in THF gave 47 (66%). Hydrogenolysis of the benzyl group in 47 afforded 16 (82%), the difluoro analogue of DAB 17, an iminosugar which has been isolated from many plant sources.

The stability of the 3-fluoro Aze motif in peptides was firmly established (Scheme 5). Reaction of the ester 24L with methylamine in methanol in the presence of calcium chloride formed the methylamide 49 (74%). Palladium-catalyzed hydrogenolysis of 49 in aqueous dioxane formed 11 (90%); unlike the corresponding 3-OH azetidine amide 6, the 3-fluoro amide 11 was stable to a wide range of pH and vulnerable to neither hydrolysis nor ring fragmentation on treatment with acid or base. A dipeptide 54 was prepared from 24L. Protection of the primary hydroxyl in 24L with TBDMS chloride in DMF in the presence of imidazole gave the fully protected silyl ether 50 (95%); hydrolysis of 50 with potassium carbonate in aqueous dioxane afforded the free acid 51. The ester 50 was converted to the corresponding methylamide 52 on reaction with methylamine in methanol in the presence of calcium chloride (100%); the benzyl group in 52 was removed by hydrogenation in the presence of palladium to give the free amine 53. The acid 51 and the amine 53 were coupled with N,N,N',N'-tetramethyl-O-(1H-benzotriazol-1-yl)uronium hexafluorophosphate (HBTU) in DMF to give the fully protected dipeptide 54 (45% from 50). The dipeptide was stable and indicated that such Aze analogues could be useful as components for peptides and for study of the predisposition of substituted Aze units to induce secondary structure.

Bioassays. Glycosidase Inhibition. The fluoro iminosugars prepared in this paper were compared with the analogous 3-OH azetidines (Figure 2) as inhibitors of the following glycosidases (Table 1, [Supporting Information\)](#page-13-0):  $37,38$  $37,38$   $\alpha$ -glucosidases (EC 3.2.1.20, rice, yeast, Aspergillus niger), β-glucosidases (EC 3.2.1.21, almond, bovine liver, A. niger),  $\alpha$ -galactosidase (EC 3.2.1.22, coffee beans),  $β$ -galactosidase (EC 3.2.1.23, bovine liver), α-mannosidase (EC





3.2.1.24, Jack bean),  $\beta$ -mannosidase (EC 3.2.1.25, snail),  $\alpha$ -Lrhamnosidase (EC 3.2.1.40, Penicillium decumbens),  $\alpha$ -L-fucosidase (EC 3.2.1.44, bovine kidney),  $\beta$ -glucuronidases (EC 3.2.1.31, Escherichia coli, bovine liver), trehalase (EC 3.2.1.28, porcine kidney), and amyloglucosidases (EC 3.2.1.3, A. niger, Rhizopus sp.),  $\beta$ -N-acetylglucosaminidases (EC 3.2.1.52, human placenta, bovine kidney, Jack bean, HL 60, Aspergillus oryzae), α-N-acetylgalactosaminidase (EC 3.2.1.49, chicken liver, Charonia lampas), and  $\beta$ -Nacetylgalactosaminidases, (EC 3.2.1.53, HL 60, A. oryzae).

Several azetidine iminosugars show specific inhibition of nonmammalian glycosidases.<sup>[24](#page-14-0)</sup> Although no significant inhibition of any glycosidase is shown either by the free acid 4 or by the iminohexitol 57, meso-azetidine triol 55 is a potent inhibitor of yeast  $\alpha$ -glucosidase (IC<sub>50</sub> 9.5  $\mu$ M) and a good inhibitor of trehalase (IC<sub>50</sub> 30  $\mu$ M) and rice  $\alpha$ -glucosidase (IC<sub>50</sub> 83  $\mu$ M).<sup>[25](#page-14-0)</sup> Potent inhibition of the yeast enzyme is rare; DAB 17 and  $\text{DMDP}$  are only natural iminosugars to do so. $^{39}$  $^{39}$  $^{39}$  The N-nonyl derivative 56 is an inhibitor of some ceramide-specific glucosyl transferases and glucosidases.[40](#page-14-0) The amide 6 is a potent inhibitor of a number of  $\beta$ -hexosaminidases but shows no inhibition of any other glycosidase; however it decomposes rapidly above pH 8.<sup>[14](#page-14-0)</sup> Assays of the fluoro azetidines 10L, 11, 14, and 15 showed there was no inhibition of any glycosidase (Table 1, [Supporting](#page-13-0) [Information\)](#page-13-0). The fluoro prolinol 16, the fluorine analogue of DAB 17, similar showed no inhibition of any glucosidase. The N-benzyl analogues 34, 35L, 42, 47, and 49 also showed no glycosidase inhibition (Table 2, [Supporting Information](#page-13-0)). Replacement of hydroxyl groups in iminosugars by fluorine substituents almost invariably reduces or eliminates glycosidase inhibition; $41$  the only exception so far reported is increased glucosidase inhibition in fluorinated australines.<sup>[42](#page-14-0)</sup>

Inhibition of Various Human Cancer Cell Growth. In the first screening, the fluoro compounds (10L, 10D, 11, 15, and 14) and their oxygen equivalents (4, 6, 55, and 57) were tested for growth inhibition effects against human pancreatic carcinoma cell line  $(PANC-1)<sup>43</sup>$  $(PANC-1)<sup>43</sup>$  $(PANC-1)<sup>43</sup>$  and were compared with 5-fluorouracil 18 and gemcitabine 19 as positive controls.<sup>[44](#page-14-0)</sup> The PANC-1 cells were incubated in an atmosphere of 5%  $CO<sub>2</sub>$  at 37 °C and subcultured every 3 days. In all experiments, cells were grown to 80−90% confluence. The growth inhibition activities were elucidated by MTT assay. PANC-1 ( $1 \times 10^4$  cells/mL) was seeded in each well of 96-well plates. After incubation with 200  $\mu$ M test compounds for 72 h, the MTT solution  $(1 \text{ mg/mL}$  in PBS: pH 7.4) was added to each well (10  $\mu$ L/well). The plate was incubated for an additional 4 h at 37 °C, and the resulting MTT-formazan crystals produced were dissolved with 200  $\mu$ L of DMSO. Absorbance was measured by FLUOstar OPTIMA (BMG LABTECH) at 540 nm. The given values were counted in triplicate. Growth inhibition was estimated as the reduction in values from a DMSO control. Preliminary results indicate that only meso-azetidine 15 showed significant inhibition of growth PANC-1 cells; different samples of 15 synthesized by the alternative pathways in Scheme [3](#page-3-0) showed the same activity. Of all the fluoroiminosugars tested, only 15 showed activity; the N-benzyl derivative 42 gave no inhibition. It is noteworthy that the fluoroazetidine diol 15 did

not inhibit any glycosidases; in contrast the meso-triol 55 was a good inhibitor of several glucosidases but had no effect on growth of cancer cells.

Thus, we next investigated the effect of the fluoro mesoazetidine 15 on growth inhibition effects against various human cancer cells (Table 1). The IC<sub>50</sub> values of *meso*-azetidine 15, fluorouracil 18, and gemcitabine 19 toward PANC-1 were  $165.3 \pm 9.1$ ,  $33.7 \pm 11.6$ , and  $122.9 \pm 66.4 \mu M$ , respectively. It is noteworthy that growth inhibition potency of 15 against PANC-1 was equivalent to gemcitabine 19. Furthermore, 15 also showed superior inhibition spectrum widely against human liver carcinoma (Hep G2), human colon adenocarcinoma (SW480), and human breast adenocarcinoma cell line (MCF-7), with IC<sub>50</sub> values of 80.3  $\pm$  6.2, 194.7  $\pm$  1.2, and 332.4  $\pm$  50.2  $\mu$ M, respectively. The inhibition mechanism of 15 is now under investigation, but the inhibition is similar to that of 18 and 19 as it had the same induction profile of apoptosis for the cancer cells and the effects depended on the incubation time.

#### ■ CONCLUSION

Because of the instability of amides of 3-OH-Aze such as 6, it is not possible to incorporate such moieties into peptides. Oxygenation of Aze-containing peptides by prolyl hydroxylases may account for the pathologies associated with human consumption of Aze. In contrast, the fluoro analogues are stable, and the enantiomeric fluoro-Aze esters 24D and 24L and the difluoroproline ester 28 provide suitable building blocks for modified azetidine and proline peptides. The azidomethyl azetidine 27 is a dipeptide isostere, which may form oligomers with disposition to form novel secondary structures. None of the fluoroazetidines show any significant inhibition of glycosidases. The inhibition of pancreatic cell growth by the mesofluoroazetidine diol 15 is a further example of biological activity of an iminosugar which is not due to glycosidase inhibition; other analogues will be studied in the hope of elucidating the mechanism.

#### **EXPERIMENTAL SECTION**

General Experimental Procedures. All commercial reagents were used as supplied. Thin-layer chromatography (TLC) was performed on aluminum sheets coated with 60  $F_{254}$  silica. Plates were visualized using a spray of 0.2% w/v cerium(IV) sulfate and 5% ammonium molybdate solution in 2 M aqueous sulfuric acid. Flash chromatography was performed on Sorbsil C60 40/60 silica. Melting points were recorded on a Kofler hot block and are uncorrected. Optical rotations are quoted in  $10^3$  deg·cm<sup>2</sup>·g<sup>-1</sup> at concentrations (c) in g·100 mL<sup>-1</sup>. <sup>1</sup>H and <sup>13</sup>C NMR spectra were assigned by utilizing 2D COSY, HSQC, and HMBC spectra. All chemical shifts  $(\delta)$  are quoted in ppm and coupling constants (J) in hertz. Residual signals from the solvents were used as an internal reference.<sup>[45](#page-14-0)</sup> For solutions in  $D_2O$  acetonitrile was used as an internal reference. HRMS measurements were made using a microTOF mass analyzer.

3-Deoxy-1,2;5,6-di-O-isopropylidene-3-fluoro-α-D-glucofuranose, 21. Triflic anhydride (11.0 mL, 38.1 mmol) was added dropwise to a solution of 1,2;5,6-di-O-isopropylidene-D-allose (5.30 g, 20.3 mmol) and anhydrous pyridine (10 mL, 76.7 mmol) in dichloromethane (25 mL) at −20 °C. After 1.5 h, TLC (cyclohexane/ethyl acetate, 1:1) indicated the consumption of the starting material  $(R_f 0.30)$  and the formation of one major product  $(R_f 0.64)$ . The reaction mixture was diluted with DCM (40 mL) and washed with HCl (2 M, aq  $3 \times 40$  mL). The organic layer was dried  $(MgSO<sub>4</sub>)$ , and the solvent was removed in vacuo to give the crude triflate  $(7.5 \text{ g})$  as yellow crystalline solid.

Cesium fluoride (9.10 g, 60.0 mmol) was added in one portion to a solution of the crude triflate (7.5 g) in 2-methyl-2-propanol (30 mL). The reaction mixture was stirred at 80 °C for 26 h until TLC (cyclohexane/ethyl acetate, 2:1) indicated consumption of the triflate  $(R_f 0.54)$  and formation of a new product  $(R_f 0.61)$ . The reaction mixture was diluted with ethyl acetate (50 mL) and washed with 1:1  $H<sub>2</sub>O/NaHCO<sub>3</sub>$  (satd aq, 50 mL) and brine (50 mL) sequentially, and the aqueous layer was back-extracted with DCM  $(3 \times 50 \text{ mL})$ . The combined organics were dried  $(MgSO<sub>4</sub>)$ , filtered, and concentrated under reduced pressure. The residue was purified by flash chromatography (cyclohexane/ethyl acetate,  $6:1 \rightarrow 4:1$ ) to give the title compound 21 as clear oil (4.70 g, 89%): HRMS (ESI +ve) found 285.1110 [M + Na]<sup>+</sup>, C<sub>12</sub>H<sub>19</sub>FNaO<sub>5</sub><sup>+</sup> requires 285.1109; [ $\alpha$ ]<sub>D</sub><sup>20</sup> –19.7 (*c* 1.06, CHCl<sub>3</sub>) [lit.<sup>[46](#page-14-0)</sup>  $[\alpha]_{\text{D}}^{20}$  –37.0 (c 1.00, CHCl<sub>3</sub>)];  $\nu_{\text{max}}$  (thin film) fingerprint region only;  $\delta_H$  (CDCl<sub>3</sub>, 400 MHz) 1.32 (3H, s, CH<sub>3</sub>), 1.36 (3H, s, CH<sub>3</sub>), 1.44  $(3H, s, CH_3)$ , 1.50  $(3H, s, CH_3)$ , 4.03  $(1H, dd, H6, J_{6,5}$  4.8,  $J_{gem}$  8.8), 4.10 (1H, ddd, H4, J<sub>4,3</sub> 2.2, J<sub>4,5</sub> 8.3, J<sub>4,F</sub> 29.1), 4.12 (1H, dd, H6', J<sub>6',5</sub> 6.1, J<sub>gen</sub> 8.8), 4.28 (1H, ddd, H5,  $J_{5,6}$  4.9,  $J_{5,6'}$  6.1,  $J_{5,4}$  8.3), 4.69 (1H, dd, H2,  $J_{2,1}$  3.8,  $J_{2,F}$  10.6), 5.01 (1H, dd, H3,  $J_{4,3}$  2.2,  $J_{3,F}$  49.9), 5.95 (1H, d, H1,  $J_{1,2}$  3.7);  $\delta$ <sub>C</sub> (CDCl<sub>3</sub>, 100 MHz) 25.1 (CH<sub>3</sub>), 26.2 (CH<sub>3</sub>), 26.7 (CH<sub>3</sub>), 27.0 (CH<sub>3</sub>), 67.2 (C6), 72.0 (d, C5,  $J_{5,F}$  7.0), 80.6 (d, C4,  $J_{4,F}$  19.1), 82.5 (d, C2,  $J_{2,F}$  33.4), 93.8 (d, C3,  $J_{3,F}$  184.4), 105.2 (C1), 109.5  $(C(CH_3)_2)$ , 112.4  $(C(CH_3)_2)$ ;  $\delta_F$  (CDCl<sub>3</sub>, 376 MHz) −207.6 (ddd,  $J_{E,2}$  10.8,  $J_{E,4}$  29.2,  $J_{F,3}$  49.8);  $m/z$  (ESI +ve) 263 ([M + H]<sup>+</sup>, 100), 285  $([M + Na]^+, 72).$ 

3-Deoxy-3-fluoro-1,2-O-isopropylidene-α-D-glucofuranose, 36. A solution of the diacetonide 21 (4.20 g, 16.0 mmol) in methanol (20 mL) and 1% aqueous sulfuric acid (20 mL) was stirred at rt for 18 h after which TLC (ethyl acetate) indicated the disappearance of starting material ( $R_f$ 0.88) and the formation of a product ( $R_f$ 0.65). The reaction mixture was neutralized with triethylamine, and the solvent was removed in vacuo to give a residue that was purified by column chromatography (cyclohexane/ethyl acetate,  $3:1 \rightarrow 0:1$ ) to form the monoacetonide 36 (3.55 g, 100%). HRMS  $m/z$  (ESI +ve) found 245.0794  $[M + Na]^+$ ,  $C_9H_{15}FNaO_5^+$  requires 245.0796;  $[\alpha]_D^2$ <sup>0</sup> -18.5 (c 0.80, CHCl<sub>3</sub>);  $\nu_{\text{max}}$  (thin film) 3411 (br, m, OH);  $\delta_{\text{H}}$  (CDCl<sub>3</sub>, 400 MHz) 1.33 (3H, s, CH3), 1.50 (3H, s, CH3), 2.68 (2H, s, OH), 3.75 (1H, ddd, H6, J 0.5,  $J_{6,5}$  5.1,  $J_{gem}$  11.5), 3.86 (1H, dd, H6',  $J_{6',5}$  3.2,  $J_{gem}$ 11.5) 3.96 (1H, ddd, H5,  $J_{5,6'}$  3.3,  $J_{5,6}$  5.3,  $J_{5,4}$  8.7), 4.16 (1H, ddd, H4,  $\check{J}_{4,3}$ 2.2,  $J_{4,5}$  8.8,  $J_{4,F}$  29.3), 4.70 (1H, dd, H2,  $J_{2,1}$  3.9,  $J_{2,F}$  10.8), 5.09 (1H, dd, H3,  $J_{3,4}$  2.2,  $J_{3,F}$  49.9), 5.96 (1H, d, H1,  $J_{1,2}$  3.7);  $\delta_C$  (CDCl<sub>3</sub>, 100 MHz) 26.2 (CH<sub>3</sub>), 26.6 (CH<sub>3</sub>), 64.1 (C6), 68.3 (d, C5, J<sub>5,F</sub> 6.4), 79.7 (d, C4, J<sub>4,F</sub> 19.1), 82.4 (d, C2,  $J_{2,F}$  31.8), 94.1 (d, C3,  $J_{3,F}$  182.8), 105.1 (C1), 112.4  $(C(CH_3)_2)$ ;  $\delta_F$  (CDCl<sub>3</sub>, 376 MHz) −208.0 (ddd, J<sub>F,2</sub> 10.8, J<sub>F,4</sub> 29.2, J<sub>F,3</sub> 49.8);  $m/z$  (ESI +ve) 245 ([M + Na]<sup>+</sup>, 100).

3-Deoxy-3-fluoro-1,2-O-isopropylidene-α-D-xylofuranose, 37. Sodium periodate (14.5 g, 67.6 mmol) was added in portions to a solution of diol 36 (12.5 g, 56.3 mmol) in 1,4-dioxane/ $H_2O$  (2:1, 60 mL). The reaction mixture was stirred at rt for 3 h until TLC (cyclohexane/ethyl acetate, 1:1) showed the consumption of starting material ( $R_f$  0.15) and the formation of a new major product ( $R_f$  0.31) after which time ethanol (50 mL) was added and stirred for a further 20 min. The white solid formed was removed by filtration, and sodium borohydride (2.13 g, 56.3 mmol) was added to the stirred reaction mixture. After 2 h, the formation of the desired product 37 and the consumption of the intermediate aldehyde was confirmed by mass spectrometry (aldehyde:  $m/z$  245  $[M + MeOH + Na<sup>+</sup>]$ ). The reaction mixture was adjusted to pH 7 by addition of acetic acid. The mixture was filtered and concentrated under reduced pressure to give a residue that was purified by column chromatography (cyclohexane/ethyl acetate,  $6:1 \rightarrow 1:1$ ) to afford the title compound 37 (9.90 g, 92%): HRMS  $m/z$ (ESI +ve) found 215.0686 [M + Na]<sup>+</sup>,  $C_8H_{13}F_{13}aO_4$ <sup>+</sup> requires 215.0690;  $[\alpha]_{\text{D}}^{20}$  –25.1 (c 0.95, CHCl<sub>3</sub>)  $[\text{lit.}^{47} [\alpha]_{\text{D}}^{20}$  $[\text{lit.}^{47} [\alpha]_{\text{D}}^{20}$  $[\text{lit.}^{47} [\alpha]_{\text{D}}^{20}$  –17.1 (c 1.06, DCM)];  $\nu_{\text{max}}$  (thin film) 3437 (br, w, OH);  $\delta_{\text{H}}$  (CDCl<sub>3</sub>, 400 MHz)

1.32 (3H, s, CH<sub>3</sub>), 1.50 (3H, s, CH<sub>3</sub>), 1.98 (1H, s, OH), 3.88 (1H, dd, H5, J<sub>5,4</sub> 5.5, J<sub>gem</sub> 11.7), 3.93 (1H, ddd, H5', J 1.2, J<sub>5',4</sub> 6.6, J<sub>gem</sub> 11.7), 4.35 (1H, dddd, H4,  $J_{4,3}$  2.3,  $J_{4,5}$  5.5,  $J_{4,5'}$  6.6,  $J_{4,F}$  30.2), 4.70 (1H, dd, H2,  $J_{2,1}$ 3.9,  $J_{2,F}$  11.2), 4.97 (1H, dd, H3,  $J_{3,4}$  2.4,  $J_{3,F}$  50.4), 5.99 (1H, d, H1,  $J_{1,2}$ 3.7);  $\delta_C$  (CDCl<sub>3</sub>, 100 MHz) 26.2 (CH<sub>3</sub>), 26.6 (CH<sub>3</sub>), 59.8 (d, C5, J<sub>5,F</sub> 9.5), 80.2 (d, C4,  $J_{4,F}$  19.1), 82.8 (d, C2,  $J_{2,F}$  33.4), 94.1 (d, C3,  $J_{3,F}$  184.4), 104.8 (C1), 112.3 ( $C(CH_3)_2$ );  $\delta_F$  (CDCl<sub>3</sub>, 376 MHz) −208.7 (ddd, J<sub>F.2</sub>) 11.4,  $J_{F,4}$  30.2,  $J_{F,3}$  50.4);  $m/z$  (ESI +ve) 215 ([M + Na]<sup>+</sup>, 100).

3-Deoxy-3-fluoro-p-xylose, 38. Dowex (50W X8, H<sup>+</sup>) (2.00 g) was added to a solution of monoacetonide 37 (9.90 g, 51.6 mmol) in 1,4-dioxane/H<sub>2</sub>O (1:1, 60 mL). The reaction mixture was stirred at 80 °C for 18 h, after which TLC analysis (ethyl acetate) indicated the disappearance of starting material  $(R_f 0.74)$  and the formation of a single product  $(R_f 0.29)$ . The reaction mixture was filtered and the solvent removed in vacuo to give a residue that was purified by column chromatography (cyclohexane/ethyl acetate,  $4:1 \rightarrow 10\%$  methanol in ethyl acetate) to give the unprotected xylose 38 (6.90 g, 88%) as a white solid in a 3:2,  $\alpha:\beta$  ratio: HRMS  $m/z$  (ESI +ve) found 175.0383 [M + Na]<sup>+</sup> .<br>ر  $C_5H_9$ FNaO<sub>4</sub><sup>+</sup> requires 175.0377; mp 106−108 °C (lit.<sup>[48](#page-14-0)</sup> mp 127 °C);  $\nu_{\text{max}}$  (thin film) 3284 (br, s, OH);  $\delta_{\text{H}}$  (CD<sub>3</sub>OD, 400 MHz) 3.20 (1H, ddd, H5 $\beta$ , J 1.2, J 10.8, J<sub>gem</sub> 11.6), 3.28–3.36 (1H, m, H2 $\beta$ ), 3.51–3.61 (2H, m, H2α, H5α), 3.65−3.78 (3H, m, H4α, H4β, H5′α), 3.84 (1H, dt, H5′ $\beta$ , J<sub>5′,4</sub> 6.1, J<sub>gem</sub> 11.5), 4.16 (1H, dt, H3 $\beta$ , J<sub>3,2</sub> = J<sub>3,4</sub> 8.8, J<sub>3,F</sub> 53.1), 4.42 (1H, d, H1 $\beta$ ,  $J_{1,2}$  7.8), 4.44 (1H, dt, H3 $\alpha$ ,  $J_{3,2} = J_{3,4}$  8.4,  $J_{3,F}$  54.4), 5.07 (1H, t, H1 $\alpha$ ,  $J_{1,2} = J_{1,F}$  3.7);  $\delta_C$  (CD<sub>3</sub>OD, 100 MHz) 61.8 (d, C5 $\alpha$ ,  $J_{5,F}$ 8.0), 65.7 (d, C5 $\beta$ , J<sub>5,F</sub> 9.5), 69.6 (d, C4 $\alpha$ , J<sub>4,F</sub> 19.1), 69.9 (d, C4 $\beta$ , J<sub>4,F</sub> 17.5), 72.2 (d, C2 $\alpha$ ,  $J_{2,F}$  15.9), 74.5 (d, C2 $\beta$ ,  $J_{2,F}$  17.5), 94.4 (d, C1 $\alpha$ ,  $J_{1,F}$ 11.1) 96.5 (d, C3 $\alpha$ ,  $J_{3,F}$  179.6) 98.2 (d, C3 $\beta$ ,  $J_{3,F}$  182.8) 98.4 (d, C1 $\beta$ ,  $J_{1,F}$ 12.7);  $\delta_F$  (CD<sub>3</sub>OD, 376 MHz) −195.8 (ddt, F $\beta$ , J 6.7, J<sub>F,2</sub> = J<sub>F,4</sub> 13.6, J<sub>F,3</sub> 52.8),  $-201.3$  to  $-201.5$  (m, Fa);  $m/z$  (ESI +ve) 175 ([M + Na]<sup>+</sup>, 100).

3-Deoxy-3-fluoro-1,2,4-tri-O-acetyl-D-xylopyranose, 39. A solution of fluoroxylose 38 (6.90 g, 45.4 mmol) in acetic anhydride/ pyridine (1:1, 60 mL) was stirred at rt for 14 h until TLC (cyclohexane/ ethyl acetate, 1:1) indicated the consumption of the starting material  $(R_f 0.00)$  and the formation of a single major product  $(R_f 0.71)$ . The solvent was concentrated under reduced pressure, and the residue was dissolved in ethyl acetate (60 mL) and washed sequentially with HCl  $(2 M, aq, 2 \times 60 \text{ mL})$ , NaHCO<sub>3</sub> (satd aq, 60 mL), and brine (60 mL). The organic layer was dried  $(MgSO<sub>4</sub>)$  and filtered, and the solvent was removed in vacuo to afford a residue which was purified by column chromatography (cyclohexane/ethyl acetate, 7:1  $\rightarrow$  5:1) to give triacetate xylopyranose 39 (11.3 g, 89%) as a white solid in an 8:5,  $\alpha$ : $\beta$  ratio of anomers: HRMS  $m/z$  (ESI +ve) found 301.0696 ([M + Na]<sup>+</sup>),  $\rm C_{11}H_{15}FNaO_7^+$  requires 301.0696; mp 44−48 °C;  $\nu_{\rm max}$  (thin film) 1751  $(s, C=O)$ ;  $\delta_H$  (CDCl<sub>3</sub>, 400 MHz) 2.09 (3H, s, CH<sub>3</sub>), 2.11 (12H, s, 4 x CH<sub>3</sub>), 2.15 (3H, s, CH<sub>3</sub>), 3.46 (1H, dd, H5 $\beta$ , J<sub>5,4</sub> 7.8, J<sub>gem</sub> 12.2), 3.61 (1H, dd, H5 $\alpha$ , J<sub>5,4</sub> 10.8, J<sub>gem</sub> 11.2), 3.97 (1H, ddd, H5' $\alpha$ , J 4.6, J 6.3, J<sub>gem</sub> 11.1), 4.18 (1H, dt, H5′ $\beta$ , J<sub>s'A</sub> = J 4.7, J<sub>gem</sub> 12.2), 4.63 (1H, dt, H3 $\beta$ , J<sub>3,2</sub> = J<sub>3,4</sub> 7.7,  $J_{3,F}$  49.9), 4.82 (1H, dt, H3 $\alpha$ ,  $J_{3,2} = J_{3,4}$  9.3,  $J_{3,F}$  53.3), 5.02–5.18 (4H, m, H2α, H2β, H4α, H4β), 5.69 (1H, d, H1β,  $J_{1,2}$  6.6), 6.27 (1H, t, H1α,  $J_{1,2} = J_{1,F}$  3.7);  $\delta_C$  (CDCl<sub>3</sub>,100 MHz) 20.5, 20.6, 20.7 (×3), 20.8 (CH<sub>3</sub>), 60.1 (d, C5 $\alpha$ , J<sub>5,F</sub> 6.4), 61.8 (d, C5 $\beta$ , J<sub>5,F</sub> 5.6), 68.2 (d, C4 $\beta$ , J<sub>4,F</sub> 21.5), 68.8 (d, C4 $\alpha$ , J<sub>4,F</sub> 17.5), 69.2 (d, C2 $\beta$ , J<sub>2,F</sub> 21.5), 69.6 (d, C2 $\alpha$ , J<sub>2,F</sub> 17.5), 88.5 (d, C3 $\alpha$ , J<sub>3,F</sub> 190.0), 89.1 (d, C3 $\beta$ , J<sub>3,F</sub> 188.4), 89.6 (d, C1 $\alpha$ , J<sub>1,F</sub> 9.5), 91.6 (d, C1 $\beta$ ,  $J_{1,F}$  8.7), 169.7, 169.1 (×2), 169.6, 169.7 (C=O);  $\delta_F$  (CDCl<sub>3</sub>, 376 MHz) −194.9 (ddt, F $\beta$ , J<sub>F,5</sub> 4.6, J<sub>F,2</sub> = J<sub>F,4</sub> 12.6, J<sub>F,3</sub> 50.4), −199.0 (dddt, F $\alpha$ , J<sub>F,1</sub> 3.4, J<sub>F,5</sub> 4.6, J<sub>F,2</sub> = J<sub>F,4</sub> 12.6, J<sub>F,3</sub> 53.1);  $m/z$  $(ESI +ve) 301 ([M + Na]<sup>+</sup>, 100).$ 

Methyl 3-Deoxy-2,4-di-O-acetyl-3-fluoro-β-D-xylopyranoside, 40. Hydrobromic acid (33% wt in acetic acid, 24.3 mL, 98.9 mmol) was added dropwise to a solution of triacetate 39 (5.50 g, 19.8 mmol) in acetic acid and DCM (7:3, 50 mL), and the reaction was stirred at 5 °C for 5 h until TLC (cyclohexane/ethyl acetate, 1:1) showed the disappearance of starting material  $(R_f 0.63)$  and the formation of a major product  $(R_f 0.77)$ . The reaction mixture was diluted with DCM (20 mL), washed successively with ice−water (50 mL), cold NaHCO<sub>3</sub> (satd aq, 50 mL), and ice−water (50 mL). The solvent was concentrated in vacuo to afford the crude bromide as an orange solid.

Silver carbonate (9.30 g, 33.4 mmol) was added to a solution of the crude bromide in methanol (90 mL), and the reaction mixture was stirred in the dark at rt for 15 h until TLC (cyclohexane/ethyl acetate, 1:1) showed the formation of major product  $(R_f 0.47)$  and no remaining starting material. After filtration, the solvent was removed in vacuo to afford a residue which was purified by column chromatography (cyclohexane/ethyl acetate,  $8:1 \rightarrow 5:1$ ) to afford the pure methyl diacetate 40 (3.00 g, 60%) as a white solid: HRMS  $m/z$  (ESI +ve) found 273.0747 [M + Na]<sup>+</sup>, C<sub>10</sub>H<sub>15</sub>FNaO<sub>6</sub><sup>+</sup> requires 273.0745; mp 78–80 °C;  $[\alpha]_D^{20}$  –71.3 (c 0.85, CHCl<sub>3</sub>) [lit.<sup>[49](#page-14-0)</sup> mp 80–81 °C;  $[\alpha]_D^{20}$  –73 (c 1.00, CHCl<sub>3</sub>)];  $\nu_{\text{max}}$  (thin film) 1749 (s, C=O);  $\delta_{\text{H}}$  (CDCl<sub>3</sub>, 400 MHz) 2.10  $(3H, s, COCH<sub>3</sub>), 2.12 (3H, s, COCH<sub>3</sub>), 3.31 (1H, dd, H5, J<sub>5.4</sub> 8.3, J<sub>ge</sub>)$ 12.0), 3.46 (3H, s, OCH<sub>3</sub>), 4.15 (1H, dt, H5',  $J_{S/4} = J_{S/F}$  4.9,  $J_{gem}$  12.0), 4.36 (1H, d, H1,  $J_{1,2}$  6.4), 4.57 (1H, dt, H3,  $J_{3,2} = J_{3,4}$  7.9,  $J_{3,F}$  50.5), 4.99– 5.08 (2H, m, H2, H4);  $\delta_C$  (CDCl<sub>3</sub>, 100 MHz) 20.7 (×2) (COCH<sub>3</sub>), 56.5 (OCH<sub>3</sub>), 60.8 (d, C5,  $J_{5,F}$  6.4), 68.9 (d, C4,  $J_{4,F}$  20.7), 70.4 (d, C2,  $J_{2,F}$  20.7), 89.7 (d, C3,  $J_{3,F}$  188.4), 101.0 (d, C1,  $J_{1,F}$  8.0), 169.2 (C=O), 169.7 (C=O);  $\delta_F$  (CDCl<sub>3</sub>, 376 MHz) −194.7 (ddt, J<sub>F,5'</sub> 5.0, J<sub>F,2</sub> = J<sub>F,4</sub> 12.9,  $J_{F,3}$  50.4);  $m/z$  (ESI +ve) 273 ([M + Na]<sup>+</sup>, 100).

Methyl 3-Deoxy-3-fluoro-β-D-xylopyranoside, 41. Sodium methoxide (177 mg, 3.3 mmol) was added to a solution of diacetate 40 (8.20 g, 32.8 mmol) in methanol (100 mL). The reaction mixture was stirred at 40 °C for 15 h when TLC analysis (cyclohexane/ethyl acetate, 1:1) indicated the disappearance of starting material  $(R_f 0.47)$  and the formation of a product  $(R_f 0.19)$ . The solvent was concentrated in vacuo and the residue purified by column chromatography (cyclohexane/ethyl acetate,  $1:1 \rightarrow 0:1$ ) to obtain the title compound 41 (5.27 g, 97%) as a white solid: HRMS  $m/z$  (ESI +ve) found 189.0527  $\begin{bmatrix} M & + N a \end{bmatrix}^+$ , ,  $C_6H_{11}FNaO_4^+$  requires 189.0534; mp 100−102 °C;  $[\alpha]_D^2$ <sup>0</sup> −61.8  $(c 0.43, \text{MeOH})$  [lit.<sup>[49](#page-14-0)</sup> mp 102−104 °C; [ $\alpha$ ]<sub>D</sub><sup>20</sup> –56.0 (c 1.00, MeOH)];  $\nu_{\text{max}}$  (thin film) 3395 (br, m, OH), 1749 (s, C=O);  $\delta_{\text{H}}$  (CD<sub>3</sub>CN, 400 MHz) 3.15 (1H, ddd, H5, J<sub>5,F</sub> 1.2, J<sub>5,4</sub> 10.2, J<sub>gem</sub> 11.6), 3.35 (1H, dddd, H2, J<sub>2,OH</sub> 4.2, J<sub>2,1</sub> 7.5, J<sub>2,3</sub> 8.7, J<sub>2,F</sub> 14.2), 3.43 (3H, s, CH<sub>3</sub>), 3.54  $(1H, d, OH, J 4.8), 3.58 (1H, d, OH, J 4.8), 3.72 (1H, ddddd, H4, J<sub>4.OH</sub>)$ 5.1,  $J_{4,5'}$  5.6,  $J_{4,3}$  8.3,  $J_{4,5}$  10.2,  $J_{4,F}$  13.4), 3.84 (1H, dt, H5',  $J_{5',4} = J_{5',F}$  5.9,  $J_{\text{gem}}$  11.6), 4.11 (1H, dd, H1, J 1.0  $J_{1,2}$  7.6), 4.17 (1H, dt, H3,  $J_{3,2} = J_{3,4}$  8.6,  $\tilde{J}_{3,F}$  52.8);  $\delta_C$  (CD<sub>3</sub>CN, 100 MHz) 57.3 (CH<sub>3</sub>), 65.2 (d, C5, J<sub>5,F</sub> 8.8), 69.0  $(d, CA, J_{4,F}$  17.5), 72.7  $(d, C2, J_{2,F}$  18.3), 97.7  $(d, C3, J_{3,F}$  182.0), 104.9  $(d,$ C1,  $J_{1,F}$  11.1);  $\delta_F$  (CD<sub>3</sub>CN, 376 MHz) −194.8 (ddt,  $J_{F,5'}$  5.7,  $J_{F,2} = J_{F,4}$ 13.7,  $j_{F,3}$  52.6);  $m/z$  (ESI +ve) 189.1 ([M + Na]<sup>+</sup>, 100).

Methyl N-Benzyl-3-fluoro-2,4-imino-2,3,4-trideoxy-β-L-ribopyranoside, 26. Triflic anhydride (14.0 mL, 125.2 mmol) was added dropwise to a solution of diol 41 (5.20 g, 31.3 mmol) and pyridine (11.0 mL, 187.8 mmol) in DCM (50 mL) at −20 °C. The reaction mixture was stirred between −20 and −10 °C for 2 h, after which TLC (cyclohexane/ethyl acetate, 1:1) showed the consumption of starting material ( $R_f$ 0.12) and the formation of one major product ( $R_f$ 0.65). The reaction mixture was diluted with DCM (30 mL) and washed with HCl  $(2 M, aq, 2 \times 70 \text{ mL})$ . The organic layer was dried  $(MgSO<sub>4</sub>)$  and filtered, and the solvent was removed in vacuo to afford the crude triflate  $(13.3 g)$ as an orange solid.

Benzylamine (13.0 mL, 156.5 mmol) was added to a solution of crude triflate (13.3 g) in acetonitrile (80 mL), and the reaction mixture was stirred at 65−70 °C for 2 h until TLC analysis (cyclohexane/ethyl acetate, 1:1) indicated the consumption of the starting material ( $R_f$ 0.65) and the formation of a single product  $(R_f 0.67)$ . The solvent was concentrated in vacuo and the residue was purified by column chromatography (cyclohexane/ethyl acetate, 7:1  $\rightarrow$  2:1) to obtain ribopyranoside 26 (7.20 g, 97%) as a light yellow oil: HRMS  $m/z$  (ESI +ve) found 238.1239  $[M + H]^+$ ,  $C_{13}H_{17}FNO_2^+$  requires 238.1238;  $[a]_D^2{}^0$  –13.7 (c 1.13, CHCl<sub>3</sub>);  $\nu_{\text{max}}$  (thin film) fingerprint region only;  $\delta_{\text{H}}$  (CDCl<sub>3</sub>, 400 MHz) 3.42 (3H, s, OCH<sub>3</sub>), 3.64 (1H, ddt, H4, J<sub>4,5</sub> = J<sub>4,5</sub><sup>,</sup> 1.7, J<sub>4,2</sub> 4.5,  $J_{4,F}$  11.0), 3.73 (1H, ddd, H2, J 1.3,  $J_{2,4}$  4.5,  $J_{2,F}$  11.6), 3.75 (1H, br-d, H5,  $J_{\text{gem}}$  11.0), 4.09 (2H, s, H6), 4.33 (1H, ddd, H5', J 1.3, J 5.1,  $J_{\text{gem}}$  10.9), 4.67 (1H, t, H1, J 1.2), 4.93 (1H, d, H3,  $J_{3,F}$  58.9), 7.15–7.37 (5H, m, ArH);  $\delta_C$  (CDCl<sub>3</sub>, 100 MHz) 51.3 (C6), 55.9 (OCH<sub>3</sub>), 61.5 (d, C5, J<sub>5,F</sub> 5.6), 64.5 (d, C4,  $J_{4,F}$  18.3), 67.4 (d, C2,  $J_{2,F}$  18.3), 93.3 (d, C3,  $J_{3,F}$  210.6), 100.2 (d, C1,  $J_{1,F}$  8.0), 126.7, 128.2, 128.3 (ArCH), 138.9 (ArC);  $\delta_F$ (CDCl3, 376 MHz) −206.5 (br-d, JF,3 59.3) m/z (ESI +ve) 238.2 ([M +  $[H]^+, 100$ ), 260 ([M + Na]<sup>+</sup>, 2).

Methyl <sup>N</sup>-Benzyl-3-fluoro-2,4-imino-2,3,4-trideoxy-L- ribonate, 24L. Ribopyranoside <sup>26</sup> (200 mg, 0.84 mmol) was dissolved

in 2 M aq HCl/1,4-dioxane (5:1, 2 mL). The reaction mixture was stirred at 40 °C for 21 h. After the consumption of starting material and the formation of aldehyde was confirmed by mass spectrometry  $(m/z)$  $(ESI +ve)$  278  $[M + MeOH + Na]<sup>+</sup>$ , the solvent was removed in vacuo to give a black solid. A solution of the black solid and potassium carbonate (349 mg, 2.53 mmol) was stirred at 0 °C under nitrogen atmosphere. Iodine solution (271 mg, 1.09 mmol), which was predissolved in anhydrous methanol (4 mL) by sonication, was added dropwise into the reaction mixture at 0  $^{\circ}{\rm C}.$  Then the mixture was stirred under 5  $^{\circ}{\rm C}$  for 2 h until the completion of reaction was confirmed by the mass spectrum. Sodium sulfite (satd aq, 5 mL) was poured into the reaction mixture, and distilled water (40 mL) was added to dissolve the precipitate. The aqueous layer was exacted with ethyl acetate  $(4 \times 50 \text{ mL})$ , and the combined organic layers were dried (MgSO<sub>4</sub>), filtrated and concentrated in vacuo to afford the crude product which was further purified by column chromatography (cyclohexane/ethyl acetate,  $5:1 \rightarrow 1:1$ ) to yield the title compound 24L as a brown oil (157 mg, 74%).

Large Scale. Ribopyranoside 26 (1.565 g, 6.6 mmol) was dissolved in 2 M aq HCl/1,4-dioxane (5:1, 120 mL). The reaction mixture was stirred at 40 °C for 21 h. After the consumption of starting material and the formation of aldehyde was confirmed by mass spectrometry  $(m/z)$  $(ESI +ve)$  278  $[M + MeOH + Na]$ <sup>+</sup>), solvent was removed in vacuo to give a black solid. A solution of the black solid and potassium carbonate  $(2.74 \text{ g}, 19.8 \text{ mmol})$  was stirred at 0 °C under nitrogen atmosphere in MeOH (60 mL). Iodine (2.18g, 8.6 mmol), which was predissolved in anhydrous methanol (60 mL) by sonication, was added dropwise into the reaction mixture at 0 °C. Then the mixture was stirred at 0 °C for 2 h until completion of reaction was confirmed by mass spectrum. Sodium sulfite (satd aq, 30 mL) was poured into the reaction mixture, and distilled water (100 mL) was added to dissolve the precipitate. The aqueous layer was exacted with ethyl acetate  $(4 \times 100 \text{ mL})$ , and the combined organic layers were dried (MgSO<sub>4</sub>), filtrated, and concentrated in vacuo to afford the crude product which was further purified by column chromatography (cyclohexane/ethyl acetate,  $5:1 \rightarrow 1:1$ ) to yield the title compound 24L as a brown oil (920 mg, 55%): HRMS  $m/z$ (ESI +ve) found 276.1014  $[M + Na]^+$ ,  $C_{13}H_{16}FNNaO_3^+$  requires 276.1006;  $[\alpha]_D^{20}$  –43.8 (c 0.54, CHCl<sub>3</sub>);  $\nu_{\text{max}}$  (thin film) 3439 (br, w, OH), 1740 (s, C=O);  $\delta_H$  (CDCl<sub>3</sub>, 400 MHz) 2.35 (1H, s, OH), 3.17 (1H, dd, H5,  $J_{5,4}$  2.7,  $J_{gem}$  12.1), 3.37 (1H, ddddd, H4, J 0.6,  $J_{4,5}$ ′ 2.2,  $J_{4,5}$ 2.7,  $J_{4,3}$  4.9,  $J_{4,F}$  21.7), 3.44 (1H, dd, H5',  $J_{5',4}$  2.1,  $J_{\text{gem}}$  12.1), 3.70 (3H, s, OCH<sub>3</sub>), 3.73 (1H, d, H6, J<sub>gem</sub> 12.5), 3.78 (1H, ddd, H2, J 0.6, J<sub>2,3</sub> 4.8, J<sub>2,F</sub> 22.2), 3.99 (1H, d, H6',  $\int_{\text{gem}}$  12.5), 5.05 (1H, dt, H3,  $J_{3,2} = J_{3,4}$  4.9,  $J_{3,F}$ 55.9), 7.26−7.36 (5H, m, ArH);  $\delta_C$  (CDCl<sub>3</sub>, 100 MHz) 52.1 (OCH<sub>3</sub>), 60.1 (d, C5,  $J_{5,F}$  4.0), 60.5 (C6), 68.0 (d, C2,  $J_{2,F}$  21.5), 69.7 (d, C4,  $J_{4,F}$ 20.7), 84.4 (d, C3,  $J_{3,F}$  213.8), 128.0, 128.6, 129.3 (ArCH), 135.9 (ArC), 170.2 (d, C1, J 5.6);  $\delta_F$  (CDCl<sub>3</sub>, 376 MHz) −181.5 (dt, J<sub>F,2</sub> = J<sub>F,4</sub> 22.0 J<sub>F,3</sub> 55.7;  $m/z$  (ESI +ve) 254.2 ([M + H]<sup>+</sup>, 100), 276.1 ([M + Na]<sup>+</sup>, 22).

Methyl N-Benzyl-3-fluoro-2,4-imino-2,3,4-trideoxy-L-ribonamide, 49. Methylamine (0.3 mL, 2.6 mmol, in absolute ethanol) was added to a solution of methyl ester 24L (32 mg, 0.13 mmol) and calcium chloride (14 mg, 0.13 mmol) in anhydrous methanol (1.5 mL). The reaction mixture was stirred at 40 °C for 2 h until the completion of reaction was confirmed by mass spectrometry  $(m/z)$  (ESI +ve) 253 [M + H]<sup>+</sup>). The reaction mixture was poured onto ethyl acetate (50 mL), dried  $(MgSO<sub>4</sub>)$ , and filtered and the solvent removed in vacuo to yield the title compound 49 as a yellow oil which was used without further purification (24 mg, 74%): HRMS m/z (ESI +ve) found 275.1166  $[M + Na]<sup>+</sup>, C<sub>13</sub>H<sub>17</sub>FN<sub>2</sub>NaO<sub>2</sub><sup>+</sup> requires 275.1166; [a]<sub>D</sub><sup>20</sup> -16.5 (c 1.22,$ CHCl<sub>3</sub>);  $\nu_{\text{max}}$  (thin film) 3337 (br, m, OH, NH), 1654 (s, C=O);  $\delta_{\text{H}}$  $(CDCl_3$ , 400 MHz) 2.62 (3H, dt, H7, J 1.2, J<sub>7,NH</sub> 4.9), 3.36–3.46 (1H, m, H4), 3.40 (1H, dd, H5, J<sub>5,4</sub> 2.8, J<sub>gem</sub> 11.9), 3.53 (1H, dd, H5', J<sub>5,4</sub> 3.1, J<sub>gem</sub> 12.1), 3.71 (1H, ddt, H2, J 1.6,  $J_{2,3}$  4.9,  $J_{2,F}$  23.2), 3.75 (1H, d, H6,  $J_{\text{gem}}$ 12.0), 3.80 (1H, d, H6′,  $J_{\text{gem}}$  12.2), 4.81 (1H, dt, H3,  $J_{3,2} = J_{3,4} = 4.6$ ,  $J_{3,5}$ 56.2), 6.70 (1H, br-s, NH), 7.26–7.35 (5H, m, ArH);  $\delta_c$  (CDCl<sub>3</sub>, 100 MHz) 25.6 (C7), 60.8 (d, C5,  $J_{5,\mathrm{F}}$  4.0), 70.3 (d, C4,  $J_{2,\mathrm{F}}$  19.1), 61.6  $(C6)$ , 70.5 (d, C2,  $J_{4,F}$  19.1), 85.8 (C3, d,  $J_{3,F}$  216.2), 128.2, 128.8, 129.3  $(ArCH)$ , 136.2  $(ArC)$ , 170.1  $(C1)$ ;  $\delta_F$   $(CDCl_3$ , 376 MHz) −177.1 (dt,  $J_{F,2} = J_{F,4}$  23.4  $J_{F,3}$  56.3);  $m/z$  (ESI +ve) 253 ([M + H]<sup>+</sup>, 100), 275 ([M +  $[Na]^{+}$ , 25).

Methyl 3-Fluoro-2,4-imino-2,3,4-trideoxy-L-ribonamide, 11. Palladium on charcoal (10% wt, 5 mg) was added to a solution of protected riboamide 49 (24 mg, 0.095 mmol) in 1,4-dioxane/ $H_2O$  (1:2, 3 mL). The reaction mixture was flushed with argon and hydrogen gas sequentially. The reaction mixture was stirred vigorously for 3 h at rt under hydrogen until the completion of reaction was confirmed by mass spectrometry  $(m/z$  (ESI +ve) 163  $[M + H]^+$ ). After filtration, the solvent was removed in vacuo to afford the title compound 11 as a light yellow oil which was used without further purification (14 mg, 90%): HRMS  $m/z$  (ESI +ve) found 185.0696 [M + Na]<sup>+</sup>, C<sub>6</sub>H<sub>11</sub>FN<sub>2</sub>NaO<sub>2</sub><sup>+</sup> requires 185.0697;  $[\alpha]_{D}^{\ \ 20}$  –61.5 (c 0.69, MeOH);  $\nu_{\rm max}$  (thin film) 3275  $\overline{b}$ (br, m, OH, NH), 1659 (s, C=O);  $\delta_H$  (CD<sub>3</sub>OD, 400 MHz) 2.81 (3H, s, H6), 3.74 (2H, s, H5), 4.28 (1H, br-d, H4,  $J_{4,F}$  16.8), 4.65 (1H, br-d, H2,  $J_{2,F}$  17.3), 5.10 (1H, br-d, H3,  $J_{3,F}$  55.5);  $\delta_C$  (CD<sub>3</sub>OD, 100 MHz) 26.0  $(C6)$ , 63.4 (d, C5,  $J_{S,F}$  4.0), 64.1 (d, C2,  $J_{2,F}$  16.0), 64.3 (d, C4,  $J_{4,F}$  16.0), 90.6 (d, C3,  $J_{3,F}$  214.0), 174.2 (C1);  $\delta_F$  (CD<sub>3</sub>OD, 376 MHz) –177.0 (dt,  $J_{F,2}$  =  $J_{F,4}$  19.8  $J_{F,3}$  54.3); m/z (ESI +ve) 163 ([M + H]<sup>+</sup>, 100), 185 ([M + Na]+ , 16).

N-Benzyl-3-fluoro-2,4-imino-2,3,4-trideoxy-L-ribonic Acid, 35L. Potassium carbonate (29 mg, 0.21 mmol) was added to a solution of methyl ester  $24L$  (40 mg, 0.16 mmol) in 1,4-dioxane/ $H_2O$  (1:2, 3 mL). The reaction mixture was stirred at 40 °C for 18 h until the completion of reaction was confirmed by mass spectrometry. HCl (2 M, aq, 0.4 mL) was added to adjust the mixture to pH 4. The solvent was removed in vacuo to obtain the crude acid, which was then loaded in 1,4 dioxane/ $H_2O$  (1:2) onto a short column of Dowex (50W X8,  $H^+$ ) (prewashed with water, 1,4-dioxane and water sequentially until the eluent was neutral). After washing again with water and 1,4-dioxane and water (1:2), the pure product was released with aqueous ammonia (2 M). Then solvent was removed in vacuo to yield the title compound 35L as a light yellow glass (22 mg, 43%): HRMS m/z (ESI +ve) found 240.1020  $[M + H]^+$ ,  $C_{12}H_{15}FNO_3^+$  requires 240.1030;  $[a]_D^{\ 20} -13.5$ (c 1.07, H<sub>2</sub>O);  $\nu_{\text{max}}$  (thin film) 3070 (br, s, OH), 1630 (s, C=O);  $\delta_{\text{H}}$  $(Py-d<sub>5</sub>, 400 MHz)$  3.63 (1H, dq, H4,  $J<sub>4,3</sub> = J<sub>4,5</sub> = J<sub>4,5</sub>$ <sup>4</sup> 4.5,  $J<sub>4,F</sub>$  22.1), 3.73 (1H, dd, H5,  $J_{5,4}$  4.6,  $J_{gem}$  11.7), 3.77 (1H, dd, H5',  $J_{5',4}$  4.0,  $J_{gem}$  11.6), 3.95  $(1H, d, H6, J_{\text{gem}} 13.0), 4.15 (1H, dd, H2, J_{2,3} 5.0, J_{2,F} 23.3), 4.35 (1H, d,$ H6′,  $J_{\text{gem}}$  13.0), 5.62 (1H, dt, H3,  $J_{3,2} = J_{3,4}$  4.9,  $J_{3,\text{F}}$  57.0), 7.22–7.64 (5H, m, ArH), 7.87 (2H, br-s, OH);  $\delta_C$  (Py- $d_5$ , 100 MHz) 61.4 (C6), 62.4 (d, C5,  $J_{5,F}$  3.2), 69.6 (d, C2,  $J_{2,F}$  18.3), 70.7 (d, C4,  $J_{4,F}$  18.3), 88.0 (d, C3,  $J_{3,F}$  210.6), 128.1, 129.0, 130.6 (ArCH), 137.8 (ArC), 174.0 (C1);  $\delta_F$  (Py-d<sub>5</sub>, 376 MHz) −178.5 (dt, J<sub>F,2</sub> = J<sub>F,4</sub> 22.8, J<sub>F,3</sub> 56.8); m/z  $(ESI +ve) 240 ([M + H]<sup>+</sup>, 100), 262 ([M + Na]<sup>+</sup>, 25).$ 

3-Fluoro-2,4-imino-2,3,4-trideoxy-L-ribonic Acid [(2R,3S,4S)- 3-Fluoro-4-(hydroxymethyl)azetidine-2-carboxylic Acid], 10L. Palladium on charcoal (10% wt, 5 mg) was added to a solution of N-benzyl-protected ribonic acid 35L (21 mg, 0.09 mmol) in 1,4 dioxane/ $H_2O$  (2 mL, 1:2). The reaction was flushed with nitrogen, argon, and hydrogen gas sequentially and stirred vigorously for 15 h at rt under hydrogen until mass spectrometry showed the completion of reaction. After filtration, the solvent was removed in vacuo to obtain a residue which was purified on a short column of  $\overline{\mathrm{Dowex}}\ (50\mathrm{W} \ \mathrm{X8}, \mathrm{H}^+ )$ (as illustrated above). The solvent was removed in vacuo to afford the title compound 10L as a light yellow glass (11 mg, 82%): HRMS  $m/z$  $(ESI -ve)$  found 148.0409 [M – H]<sup>-</sup>, C<sub>5</sub>H<sub>7</sub>FNO<sub>3</sub><sup>-</sup> requires 148.0415;  $[\alpha]_{D}^{20}$  –30.6 (c 0.50, H<sub>2</sub>O);  $\nu_{max}$  (thin film) 3233 (br, s, OH), 1630 (s, C=O);  $\delta_H$  (D<sub>2</sub>O, 400 MHz) 3.91 (1H, dd, H5, J<sub>5,4</sub> 3.7, J<sub>gem</sub> 13.2), 3.97  $(1H, dd, H5', J<sub>5',4</sub> 3.9, J<sub>gem</sub> 13.2), 4.66 (1H, dq, H4, J<sub>4,3</sub> = J<sub>4,5</sub> = J<sub>4,5'</sub> 4.2, J<sub>4,F</sub>$ 19.3), 4.92 (1H, dd, H2,  $J_{2,3}$  4.8,  $J_{2,F}$  21.3), 5.31 (1H, dt, H3,  $J_{3,2} = J_{3,4}$  4.8,  $J_{3,F}$  56.1);  $\delta_C$  (D<sub>2</sub>O, 100 MHz) 58.0 (d, C5,  $J_{5,F}$  4.0), 63.8 (d, C2,  $J_{2,F}$ 24.6), 64.9 (d, C4,  $J_{4,F}$  26.2), 87.9 (d, C3,  $J_{3,F}$  210.6), 170.4 (C1);  $\delta_F$  $(D_2O, 376 MHz) -178.3 (dt, J_{F,2} = J_{F,4} 20.4, J_{F,3} 56.1); m/z (ESI -ve):$ 148 ([M – H]<sup>+</sup>, 100).

N-Benzyl-3-fluoro-2,4-imino-2,3,4-trideoxy-meso-ribitol, 42. Method 1. The bicyclic azetidine 26 (100 mg, 0.42 mmol) was dissolved in 2 M aq HCl/1,4-dioxane (5:1, 1 mL). The reaction mixture was stirred at 40 °C for 23 h after which the consumption of starting material and the formation of aldehyde were confirmed by mass spectrometry  $(m/z$  (ESI +ve) 278 [M + MeOH + Na]<sup>+</sup>). The solvent was removed in vacuo to give a black glass.

Sodium borohydride (196 mg, 2.52 mmol) was added to a solution of the black residue in methanol (4 mL). After the solution was stirred at rt for 2 h, mass spectrometry  $(m/z)$  248) showed the completion of reaction. The solvent was concentrated in vacuo to obtain a residue which was purified on a short column of Dowex  $(50W X8, H<sup>+</sup>)$ (prewashed with water, 1,4-dioxane, and water sequentially until the eluent was neutral) to yield the desired diol 42 as a brown oil (39 mg, 40% over two steps).

Method 2. Sodium borohydride (15 mg, 0.39 mmol) was added into a solution of methyl ester 24L (100 mg, 0.39 mmol) in methanol (2 mL) at 0 °C. After the solution was stirred at rt for 3 h, mass spectrometry  $(m/z 248)$  showed the completion of reaction. The solvent was removed in vacuo to obtain a residue that was purified on a short column of Dowex (50W X8, H+ ) (prewashed with water, 1,4-dioxane, and water sequentially until the eluent was neutral) to yield the desired diol 42 as a brown oil (79 mg, 90%): HRMS m/z (ESI +ve) found 248.1057 [M +  $\text{Na}$ ]<sup>+</sup>, C<sub>12</sub>H<sub>16</sub>FNNaO<sub>2</sub><sup>+</sup> requires 248.1057; [ $\alpha$ ]<sub>D</sub><sup>20</sup> 0.0 ( $\epsilon$  1.0, CH<sub>3</sub>OH);  $\nu_{\text{max}}$  (thin film) 3355 (br, s, OH);  $\delta_{\text{H}}$  (CDCl<sub>3</sub>, 400 MHz) 2.81 (2H, br-s, OH x 2), 3.25 (2H, ddt, H2(4),  $J_{2,1} = J_{2,1'}$  3.2,  $J_{2,3}$  5.1,  $J_{2,F}$  22.8), 3.36 (2H, dd, H1(5), J<sub>1,2</sub> 3.2, J<sub>gem</sub> 11.7), 3.52 (2H, dd, H1'(5'), J<sub>1',2</sub> 2.8, J<sub>gem</sub> 11.9), 3.78 (2H, s, H6), 4.93 (1H, dt, H3,  $J_{3,2} = J_{3,4}$  4.7,  $J_{3,F}$  56.8), 7.27–7.36  $(SH, m, ArH); \delta_C (CDCl_3, 100 MHz) 60.8 (d, C1(5), J4.8), 61.2 (C6),$ 70.3 (d, C2(4), J 19.9), 83.9 (d, C3, J 207.4), 127.9, 128.6, 129.1 (ArCH), 137.0 (ArC);  $\delta_F$  (CDCl<sub>3</sub>, 376 MHz) −184.2 (dt, J<sub>F,2</sub> = J<sub>F,4</sub> 22.9,  $J_{F,3}$  56.1);  $m/z$  (ESI +ve) 248 ([M + Na]<sup>+</sup>, 100).

3-Fluoro-2,4-imino-2,3,4-trideoxy-meso-ribitol, 15. Palladium on charcoal (10% wt, 5 mg) was added to a solution of N-benzylprotected ribonic acid 42 (20 mg, 0.09 mmol) in 1,4-dioxane/H<sub>2</sub>O (1:2). The reaction was flushed with nitrogen, argon, and hydrogen gas sequentially and stirred vigorously for 5 h at rt under hydrogen until mass spectrometry showed the completion of the reaction  $(m/z)$  (ESI) +ve)  $136$  [M + H]<sup>+</sup>). After filtration, the solvent was concentrated in vacuo to obtain a residue which was purified by a short column of Dowex (50W X8, H+ ) (as illustrated above). The solvent was concentrated in vacuo to afford the title compound 15 as a light yellow oil (12 mg, 100%): HRMS  $m/z$  (ESI +ve) found 158.0590  $[M + Na]^{T}$ , ,  $C_5H_{10}$ FNNa $O_2^+$  requires 158.0588;  $\left[\alpha\right]_D{}^{20}$  0.0 (c 1.0, CH<sub>3</sub>OH);  $\nu_{\text{max}}$ (thin film) 3330 (br, s, OH);  $\delta_{\rm H}$  (D<sub>2</sub>O, 400 MHz) 3.66 (4H, d, H1(5),  $J_{1,2}$  5.7), 3.90 (2H, dq, H2(4),  $J_{2,1} = J_{2,3} = J$  5.8,  $J_{2,F}$  21.4), 4.80 (1H, dt, H3,  $J_{3,2} = J_{3,4}$  5.7,  $J_{3,F}$  56.0);  $\delta_C$  (D<sub>2</sub>O, 100 MHz) 62.3 (d, C2(4),  $J_{2,F}$ 20.0), 62.8 (d, C1(5),  $J_{1,F}$  3.8), 89.3 (d, C3,  $J_{3,F}$  209.8);  $\delta_F$  (D<sub>2</sub>O, 376 MHz) −174.8 (dt,  $J_{F,2} = J_{F,4}$  21.8,  $J_{F,3}$  56.4); m/z (ESI +ve) 136  $([M + H]^+, 100).$ 

Methyl N-Benzyl-3-fluoro-2,4-imino-5-O-mesyl-2,3,4-trideoxy-L-ribonate, 43. Methanesulfonyl chloride (0.05 mL, 0.60 mmol) was added to a solution of methyl ester 24L (100 mg, 0.40 mmol) in pyridine (3 mL) at 0 °C. The reaction mixture was stirred at 0 °C for 1 h when TLC (cyclohexane/ethyl acetate, 1:1) showed the consumption of starting material  $(R_f 0.38)$  and the formation of product  $(R_f 0.46)$ . The solvent was removed in vacuo, and the residue was purified by flash column chromatography (cyclohexane/ethyl acetate,  $5:1 \rightarrow 2:1$ ) to give the product 43 as a yellow oil (133 g, 100%): HRMS  $m/z$  (ESI +ve) found 354.0777 [M + Na]<sup>+</sup>, C<sub>14</sub>H<sub>18</sub>FNNaO<sub>5</sub>S<sup>+</sup> requires 354.0782;  $[\alpha]_{\text{D}}^{20}$  -22.1 (c 0.57, CHCl<sub>3</sub>);  $\nu_{\text{max}}$  (thin film) 1744 (s, C=O);  $\delta_H$  (CDCl<sub>3</sub>, 400 MHz) 2.97 (3H, s, CH<sub>3</sub>), 3.44 (1H, dq, H4, J<sub>4,3</sub>  $= J_{4,5} = J_{4,5'}$  4.6,  $J_{4,F}$  20.7), 3.66 (3H, s, CH<sub>3</sub>), 3.73 (1H, dd, H2,  $J_{2,3}$  5.1,  $J_{2,\mathrm{F}}$  21.3), 3.79 (1H, d, H6,  $J_{gem}$  12.7), 3.93 (1H, d, H6′,  $J_{gem}$  12.5), 3.97  $(1H, dd, H5, J<sub>5,4</sub> 4.2, J<sub>gem</sub> 11.3), 4.10 (1H, dd, H5', J<sub>5',4</sub> 4.5, J<sub>gem</sub> 11.6), 4.95)$ (1H, dt, H3,  $J_{3,2} = \tilde{J}_{3,4}$  5.0,  $J_{3,\bar{F}}$  55.3), 7.25−7.36 (5H, m, ArH);  $\delta_c$  $(CDCl_3, 100 MHz)$  37.6  $(SO_2CH_3)$ , 52.1  $(CCH_3)$ , 60.5  $(C6)$ , 66.4 (d, C4,  $J_{4,F}$  21.5), 67.7 (d, C2,  $J_{2,F}$  21.5), 67.9 (d, C5,  $J_{5,F}$  4.0), 84.5 (d, C3,  $J_{3,F}$ 217.0), 128.0, 128.5, 129.6 (ArCH), 135.3 (ArC), 169.8 (C1, d, J 4.8);  $\delta_F$  $(CDCl_3$ , 376 MHz) −179.6 (dt, J<sub>F,2</sub> = J<sub>F,4</sub> 20.9, J<sub>F,3</sub> 55.3); m/z (ESI +ve)  $332 ([M + H]<sup>+</sup>, 100).$ 

Methyl N-Benzyl-5-azido-3-fluoro-2,4-imino-2,3,4,5-tetradeoxy-L-ribonate, 27. Sodium azide (13 mg, 0.20 mmol) was added to a solution of mesylate 43 (50 mg, 0.15 mmol) in DMF (2 mL), and the reaction mixture was stirred at 60 °C for 26 h. After this time, TLC analysis (cyclohexane/ethyl acetate, 1:1) indicated the disappearance of the starting material ( $R_f$  0.48) and the formation of a single product

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 $(R_f 0.78)$ . The reaction mixture was diluted with ethyl acetate (20 mL) and washed with 1:1 H<sub>2</sub>O/brine (satd aq)  $(3 \times 20 \text{ mL})$ . The organic layer was dried (MgSO<sub>4</sub>) and filtered, and the solvent was removed in vacuo to obtain a residue that was purified by flash column chromatography (cyclohexane/ethyl acetate,  $7:1 \rightarrow 5:1$ ) to yield the title compound 27 as a light yellow oil (32 mg, 80%): HRMS  $m/z$  (ESI +ve) found 301.1072  $[M + Na]^+$ ,  $C_{13}H_{15}FN_4NaO_2^+$  requires 301.1071;  $[\alpha]_{\text{D}}^{20}$  –61.7 (c 1.62, CHCl<sub>3</sub>);  $\nu_{\text{max}}$  (thin film) 2102 (s, N<sub>3</sub>), 1743 (s, C=O);  $\delta_H$  (CDCl<sub>3</sub>, 400 MHz) 3.05 (1H, dd, H5, J<sub>5,4</sub> 4.4, J<sub>gem</sub> 13.2), 3.10 (1H, dd, H5',  $J_{5',4}$  4.4,  $J_{\text{gem}}$  13.2), 3.33 (1H, ddq, H4, J 0.7,  $J_{4,3} = J_{4,5} = J_{4,5'}$ 4.7,  $J_{4,F}$  21.2), 3.70 (3H, s, CH<sub>3</sub>), 3.70 (1H, ddd, H2, J 0.7,  $J_{2,3}$  5.0,  $J_{2,F}$ 21.5), 3.75 (1H, d, H6, J<sub>gem</sub> 12.5), 4.00 (1H, d, H6', J<sub>gem</sub> 12.7), 4.96 (1H, dt, H3,  $J_{3,2} = J_{3,4}$  5.0,  $J_{3,\overline{F}}$  55.7), 7.30–7.35 (5H, m, ArH);  $\delta_C$  (CDCl<sub>3</sub>,  $100 \text{ MHz}$ )  $51.5$  (d, C5,  $J_{5,F}$  4.0), 52.1 (CH<sub>3</sub>), 60.9 (C6), 67.4 (d, C4,  $J_{4,F}$ 20.7), 68.0 (d, C2,  $J_{2,F}$  21.5), 85.3 (d, C3,  $J_{3,F}$  215.4), 127.9, 128.5, 129.7 (ArCH), 135.6 (ArC), 170.0 (d, C1,  $J_{1,F}$  5.6);  $\delta_F$  (CDCl<sub>3</sub>, 376 MHz)  $-179.7$  (dt,  $J_{F,2} = J_{F,4}$  21.5,  $J_{F,3}$  55.7);  $m/z$  (ESI +ve) 301 ([M + Na]<sup>+</sup>, , 100).

Methyl N-Benzyl-5-O-(tert-butyldimethylsilyl)-3-fluoro-2,4 imino-2,3,4-trideoxy-L-ribonate, 50. tert-Butyldimethylsilyl chloride (111 mg, 0.74 mmol) was added to a solution of methyl ester 24L (157 mg, 0.62 mmol) in DMF (10 mL), and the reaction mixture was stirred at rt for 2 h. After this time, TLC analysis (cyclohexane/ethyl acetate, 1:1) indicated the disappearance of starting material  $(R_f 0.50)$ and the formation of a major product  $(R_f 0.87)$ . The reaction mixture was diluted with ethyl acetate (20 mL) and washed with 1:1  $H_2O/b$ rine (satd aq)  $(2 \times 20 \text{ mL})$ . The organic layer was dried  $(MgSO_4)$  and filtered, and the solvent was removed in vacuo to yield the title compound 50 as a brown oil (218 mg, 95%): HRMS  $m/z$  (ESI +ve) found 390.1878  $[M + Na]$ <sup>+</sup>, C<sub>19</sub>H<sub>30</sub>FNNaO<sub>3</sub>Si<sup>+</sup> requires 390.1871;  $[\alpha]_D^2$ <sup>20</sup> −16.4 (c 0.90, CHCl<sub>3</sub>);  $\nu_{\text{max}}$  (thin film) 1745 (s, C=O);  $\delta_{\text{H}}$  (CDCl<sub>3</sub>, 400 MHz) 0.00, 0.01 (2  $\times$  3H, 2s, CH<sub>3</sub>), 0.87 (9H, s, C(CH<sub>3</sub>)<sub>3</sub>), 3.27 (1H, dq, H4,  $J_{4,3} = J_{4,5} = J_{4,5'}$  5.4,  $J_{4,F}$  21.5), 3.47 (1H, ddd, H5, J 1.0,  $J_{5,4}$ 5.4,  $J_{\text{gem}}$  10.8), 3.51 (1H, dd, H5',  $J_{\text{S}'$ , 45.7,  $J_{\text{gem}}$  10.8), 3.64 (1H, dd, H2,  $J_{2,3}$ ) 5.1,  $J_{2,F}$  21.5), 3.65 (3H, s, OCH<sub>3</sub>), 3.82 (1H, d, H6,  $J_{\text{gem}}$  12.9), 3.91 (1H, d, H6′,  $J_{\text{gem}}$  12.9), 4.85 (1H, dt, H3,  $J_{3,2} = J_{3,4}$  5.1,  $J_{3,F}$  56.2), 7.25–7.35 (5H, m, ArH);  $\delta_C$  (CDCl<sub>3</sub>, 100 MHz) -5.5, -5.4 (CH<sub>3</sub>Si), 18.2  $(C(CH<sub>3</sub>)<sub>2</sub>), 25.8 (C(CH<sub>3</sub>)<sub>2</sub>), 51.9 (OCH<sub>3</sub>), 60.9 (C6), 63.7 (d, C5, J<sub>5,F</sub>)$ 4.0), 67.8 (d, C2,  $J_{2,F}$  21.5), 69.3 (d, C4,  $J_{4,F}$  19.1), 86.2 (d, C3,  $J_{3,F}$  214.6), 127.6, 128.3, 129.7 (ArCH), 136.0 (ArC), 170.6 (d, C1,  $J_{1,F}$  4.8);  $\delta_F$  $(CDCl_3$ , 376 MHz) −179.0 (dt, J<sub>F,2</sub> = J<sub>F,4</sub> 21.8, J<sub>F,3</sub> 56.1); m/z (ESI +ve)  $368 ([M + H]^{+}, 100), 390 ([M + Na]^{+}, 30).$ 

Methyl N-Benzyl-5-O-(tert-butyldimethylsilyl)-3-fluoro-2,4 imino-2,3,4-trideoxy-L-ribonamide, 52. Methylamine (0.30 mL, 2.6 mmol, in absolute ethanol) was added to a solution of methyl ester 50 (42 mg, 0.11 mmol) and calcium chloride (12.0 mg, 0.11 mmol) in anhydrous methanol (1.0 mL). The reaction mixture was stirred at 45 °C for 2 h when the completion of reaction was confirmed by mass spectrometry  $(m/z \text{ (ESI +ve) } 367 \text{ [M + H]}^+)$ . The pH of the reaction mixture was adjusted to pH 5 using NH<sub>4</sub>Cl (satd aq)/H<sub>2</sub>O (1:3, 2 mL), and the mixture was extracted with ethyl acetate  $(3 \times 15 \text{ mL})$ . The organic layer was dried  $(MgSO<sub>4</sub>)$  and filtered, and solvent was removed in vacuo to yield the amide 52 as a yellow oil (41 mg, 100%): HRMS  $m/z$ (ESI +ve) found 389.2037 [M + Na]<sup>+</sup>, C<sub>19</sub>H<sub>31</sub>FN<sub>2</sub>NaO<sub>2</sub>Si<sup>+</sup> requires 389.2031;  $[\alpha]_D^{20}$  –4.4 (c 0.82, CHCl<sub>3</sub>);  $\nu_{\text{max}}$  (thin film) 1681 (s, C= O);  $\delta_{\rm H}$  (CDCl<sub>3</sub>, 400 MHz) 0.01 (6H, s, CH<sub>3</sub>), 0.88 (9H, s, C(CH<sub>3</sub>)<sub>3</sub>), 2.62 (3H, d, NCH<sub>3</sub>, J<sub>CH3,NH</sub> 4.9), 3.32−3.42 (1H, m, H4), 3.38 (1H, dd, H5, J 3.3, J<sub>gem</sub> 11.1), 3.53 (1H, dd, H5', J<sub>5',4</sub> 3.5, J<sub>gem</sub> 10.9), 3.68 (1H, dd, H2,  $J_{2,3}$  4.5,  $J_{2,F}$  24.1), 3.73 (1H, d, H6,  $J_{gem}$  12.5), 3.80 (1H, d, H6',  $J_{gem}$ 12.2), 4.73 (1H, dt, H3,  $J_{3,2} = J_{3,4}$  4.4,  $J_{3,F}$  56.2), 6.88 (1H, q, NH,  $J_{NH,CH3}$ 5.0), 7.25−7.35 (5H, m, ArH);  $\delta_C$  (CDCl<sub>3</sub>, 100 MHz) −5.6, −5.5 (CH<sub>3</sub>Si), 18.2 (C(CH<sub>3</sub>)<sub>2</sub>), 25.4 (NCH<sub>3</sub>), 25.7 (C(CH<sub>3</sub>)<sub>2</sub>), 61.7 (C6), 62.2 (d, C5,  $J_{\text{S,F}}$  4.8), 70.4 (d, C4,  $J_{\text{4,F}}$  19.9), 70.5 (d, C2,  $J_{\text{2,F}}$  20.7), 86.2  $(d, C3, J_{3,F} 216.2), 127.9, 128.6, 129.2 (ArCH), 136.6 (ArC), 170.4 (d,$ C1,  $J_{1,F}$  5.6);  $\delta_F$  (CDCl<sub>3</sub>, 376 MHz) –176.3 (dt,  $J_{F,2} = J_{F,4}$  24.3,  $J_{F,3}$  56.4);  $m/z$  (ESI +ve) 367 ([M + H]<sup>+</sup>, 100), 389 ([M + Na]<sup>+</sup>, 30).

Methyl N-(N-Benzyl-8-fluoro-7,9-imino-8,7,9-trideoxy-10-O- (tert-butyldimethylsilyl)-L-ribonamido)-3-fluoro-2,4-imino-2,3,4-trideoxy-5-O-(tert-butyldimethylsilyl)-L-ribonamide, 54. Potassium carbonate (19 mg, 0.14 mmol) was added to a solution of methyl ester 50 (40 mg, 0.11 mmol) in 1,4-dioxane/ $H_2O$  (2 mL, 1:1). The reaction mixture was stirred at 40 °C for 26 h until mass spectrometry indicated completion of the hydrolysis  $(m/z \text{ (ESI -ve)}$ : 352  $[M - H]$ <sup>-</sup>), and the solvent was removed in vacuo.

Palladium on charcoal (10% wt., 5 mg) was added to a solution of 52  $(40 \text{ mg}, 0.11 \text{ mmol})$  in 1,4-dioxane/H<sub>2</sub>O  $(3 \text{ mL}, 1:2)$ . The reaction was flushed with argon and hydrogen gas sequentially and then stirred vigorously for 5 h at rt under hydrogen until mass spectrometry showed the completion of reaction  $(m/z$  (ESI +ve) 277  $[M + H]^+$ ). After filtration, the solvent was removed in vacuo to afford a residue that was used without further purification.

N,N,N′,N′-Tetramethyl-O-(1H-benzotriazol-1-yl)uronium hexafluorophosphate (HBTU, 50 mg, 0.13 mmol) was added to a solution of the crude acid (57 mg) and amine (30 mg) in anhydrous DMF (1.5 mL). After the mixture was stirred for 20 min, triethylamine (0.02 mL) was added to the reaction mixture, which was stirred rt for a further 20 h until TLC analysis (cyclohexane/ethyl acetate, 1:1) showed the consumption of the starting materials and formation of one major product  $(R_f 0.75)$ . The reaction mixture was diluted with ethyl acetate (20 mL) and washed with half saturated brine (20 mL). The organic layer was dried  $(MgSO_4)$  and filtered and the solvent removed in vacuo to give a residue that was purified by flash column chromatography (cyclohexane/ethyl acetate, 1:1) to give the pure peptide 54 as a yellow oil (30 mg, 45%): HRMS  $m/z$  (ESI +ve) found 634.3288 [M + Na]<sup>+</sup>, ,  $C_{30}H_{51}F_2N_3NaO_4Si_2^+$  requires 634.3278;  $[\alpha]_D^2$ <sup>20</sup> –44.0 (c 0.93, CHCl<sub>3</sub>);  $\nu_{\text{max}}$  (thin film) 1682 (s, C=O);  $\delta_{\text{H}}$  (CDCl<sub>3</sub>, 400 MHz) 0.01, 0.03, 0.06, 0.07 (12H, 4s, CH<sub>3</sub>), 0.85 (9H, s, C(CH<sub>3</sub>)<sub>3</sub>), 0.88 (9H, s, C(CH<sub>3</sub>)<sub>3</sub>), 2.74 (3H, d, NCH<sub>3</sub>,  $J_{CH3,NH}$  4.9), 3.36 (1H, dq, H9,  $J_{9,8} = J_{9,10} = J_{9,10'}$  5.0,  $J_{9,F}$  21.9), 3.56 (1H, dd, H7,  $J_{7,8}$  4.9,  $J_{7,F}$  20.3), 3.50–3.64 (1H, m, H4), 3.57−3.63 (1H, m, H5), 3.70 (2H, br-s, -CH<sub>2</sub>Ar), 3.74 (2H, br-s, H10), 4.19 (1H, d, H5',  $J_{\rm gem}$ 12.2), 4.31 (1H, br-d, H2,  $J_{\rm 2,F}$ 23.7), 4.79 (1H, dt, H8,  $J_{8,7} = J_{8,9}$  4.8,  $J_{8,F}$  56.1), 5.08 (1H, br-d, H3,  $J_{3,F}$  55.3), 7.26–7.35  $(SH, m, ArH)$ , 7.43 (1H, br-s, NH);  $\delta_C$  (CDCl<sub>3</sub>, 100 MHz) –5.7, –5.4  $(x 2)$  (CH<sub>3</sub>Si), 18.2 (C(CH<sub>3</sub>)<sub>2</sub>), 25.7 (NHCH<sub>3</sub>), 25.8 (C(CH<sub>3</sub>)<sub>3</sub>), 62.5  $(C5, -CH<sub>2</sub>Ar)$ , 64.1  $(C10)$ , 66.0  $(C7)$ , 67.0  $(C2)$ , 68.6  $(d, CA, J<sub>4,F</sub> 25.4)$ , 70.9 (d, C9,  $J_{9,F}$  18.3), 85.5 (d, C3,  $J_{3,F}$  194.7), 86.1 (d, C8,  $J_{8,F}$  218.6), 128.3, 128.6, 128.9 (ArCH), 136.2 (ArC), 167.7, 167.8 (C1, C6);  $\delta_F$ (CDCl<sub>3</sub>, 376 MHz) −177.7 (dt, J<sub>F,7</sub> = J<sub>F,9</sub> 21.2, J<sub>F,8</sub> 56.1), −184.4 (dt,  $J_{\text{F,2}}$  = J<sub>F,4</sub> 22.9, J<sub>F,3</sub> 55.3); m/z (ESI +ve) 612 ([M + H]<sup>+</sup>, 100), 634 ([M + Na]<sup>+</sup>, 30).

N-Benzyl-3R,4R-difluoro-L-proline Methyl Ester, 28. A solution of azetidine methyl ester 24L (289 mg, 0.79 mmol) in DCM (2 mL) was added dropwise to a solution of XtalFluor-M (289 mg, 1.19 mmol) and TEA·3HF (0.26 mL, 1.19 mmol) in DCM (2 mL) at −78 °C. After being stirred for 1 h, the mixture was stirred at rt for 18 h until TLC (cyclohexane/ethyl acetate, 2:1) showed the disappearance of starting material ( $R_f$  0.16) and the formation of major product ( $R_f$  0.56). The mixture was diluted with half saturated  $\text{NaHCO}_3 \left( \text{10 mL} \right)$  and stirred for a further 20 min before extraction with DCM  $(3 \times 20 \text{ mL})$ . The combined organic layers were dried (MgSO4), filtered, and concentrated under reduced pressure to obtain a crude residue which was purified by flash chromatography (cyclohexane/ethyl acetate,  $7:1 \rightarrow 6:1$ ) to yield the desired product 28 as a yellow oil (169 mg, 84%): HRMS  $m/z$  (ESI +ve) found 278.0958  $[M + Na]^+$ ,  $C_{13}H_{15}F_2NNaO_2^+$  requires 278.0963;  $[\alpha]_{D}^{20}$  –64.2 (c 0.31, CHCl<sub>3</sub>);  $\nu_{\text{max}}$  (thin film) 1746 (s, C=O);  $\delta_{H}$  $(CDCl_3$ , 400 MHz) 2.85 (1H, dddd, H5,  $J_{5,3F}$  2.0,  $J_{5,4}$  5.0,  $J_{gem}$  11.7,  $J_{5,4F}$ 31.9), 3.26 (1H, ddt, H5',  $J_{5',3} = J_{5',4}$  1.4,  $J_{\text{gem}}$  11.6,  $J_{5',4F}$  20.9), 3.51 (1H, dd, H2,  $J_{2,3}$  3.7,  $J_{2,3F}$  26.9), 3.66 (1H, d, H6,  $J_{\text{gem}}$  13.1), 3.76 (3H, s, OCH<sub>3</sub>), 4.05 (1H, d, H6',  $J_{\text{gem}}$  13.2), 5.05 (1H, dddt, H4,  $J_{4,3} = J_{4,5'}$  1.3,  $J_{4,5}$  4.9,  $J_{4,3\mathrm{F}}$  14.9,  $J_{4,4\mathrm{F}}$  50.7), 5.27 (1H, dddt, H3,  $J_{3,4}$  =  $J_{3,5^{\prime}}$  1.3,  $J_{3,2}$  3.7,  $J_{3,4F}$  16.6,  $J_{3,3F}$  50.1), 7.25–7.36 (5H, m, ArH);  $\delta_C$  (CDCl<sub>3</sub>, 100 MHz) 52.4 (OCH<sub>3</sub>), 56.2 (dd, C5, J<sub>5,3F</sub> 2.4, J<sub>5,4F</sub> 23.1), 57.5 (C6), 69.8 (dd, C2,  $J_{2,3F}$  0.8,  $J_{2,4F}$  26.2), 97.9 (dd, C4,  $J_{4,3F}$  29.0,  $J_{4,4F}$  182.4), 97.9 (dd, C3,  $J_{3,4F}$ 32.7,  $J_{3,3F}$  186.4), 127.5, 128.4, 128.9 (ArCH), 136.7 (ArC), 170.4 (d, C1,  $J_{1,F}$  8.8);  $\delta_F$  (CDCl<sub>3</sub>, 376 MHz) −184.6 (ddddd, 3F,  $J_{3F,5}$  2.3,  $J_{3F,4F}$ 8.0, J3F,4 14.9, J3F,2 26.3, J3F,3 50.4), −186.6 (ddddd, 4F, J4F,3F 7.9, J4F,3 16.4,  $J_{\rm 4F,5^{\prime}}$  20.9,  $J_{\rm 4F,5}$  32.0,  $J_{\rm 4F,4}$  50.7);  $m/z$  (ESI +ve) 256 ([M + H]<sup>+</sup>, 100%).

<sup>N</sup>-Benzyl-2R,3R-difluoro-2,4-imino-1,2,3,4-tetradeoxyl-D- arabinofuranose, 47. Lithium aluminum hydride (1 M in THF) (0.24 mL, 0.24 mmol) was added dropwise to a solution of difluoromethyl

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ester 28 (30 mg, 0.120 mmol) in anhydrous THF at−78 °C. The reaction mixture was stirred for 1 h until TLC analysis (cyclohexane/ethyl acetate, 2:1) showed the consumption of the starting material  $(R_f 0.67)$  and formation of a major product  $(R_f 0.37)$ . The reaction was quenched with  $NH<sub>4</sub>Cl$  (satd aq) to pH 6 and extracted with ethyl acetate (2  $\times$  15 mL). The organic layers were dried  $(MgSO<sub>4</sub>)$ , filtered, and concentrated in vacuo to yield pure alcohol 47 as a clear oil which was used without further purification (18 mg, 66%): HRMS  $m/z$  (ESI +ve) found 250.1008  $[M + Na]<sup>+</sup>$ , C<sub>12</sub>H<sub>15</sub>F<sub>2</sub>NNaO<sup>+</sup> requires 250.1014;  $[\alpha]_D^{20}$  –70.6 (c 0.90, MeOH);  $\nu_{\text{max}}$  (thin film) 3348 (br, w, OH);  $\delta_{\text{H}}$  (CD<sub>3</sub>OD, 400 MHz) 2.72 (1H, dddd, H1,  $J_{1,3F}$  3.0,  $J_{1,2}$  4.6,  $J_{gem}$  12.1,  $J_{1,2F}$  36.7), 2.74–2.85 (1H, m, H4), 3.09 (1H, ddq, H1',  $J = J = \tilde{J}$  1.0,  $J_{\text{gem}}$  12.1,  $J_{\text{1}'\text{,2F}}$  20.5), 3.48 (1H, d, H6, J $_{\mathit{gem}}$  13.2), 3.64 (1H, ddt, H5, J = J 0.7, J $_{5,4}$  6.1, J $_{\mathit{gem}}$  11.5), 3.67 (1H, ddt,  $\text{H5}^{\prime}$ , J = J 1.1, J<sub>5',4</sub> 4.9, J<sub>gem</sub> 11.6), 4.05 (1H, d, H6', J<sub>gem</sub> 13.1), 4.91–5.11  $(2H, m, H3, H2), 7.23–7.36 (5H, m, ArH); \delta_C (CD_3OD, 100 MHz) 58.3$  $(dd, C1, J_{1,3F}$  2.4,  $J_{1,2F}$  22.3), 59.5 (C6), 61.6 (d, C5,  $J_{5,4F}$  6.4), 71.3 (dd, C4, J 1.6,  $J_{4.3F}$  22.3), 95.3 (dd, C2,  $J_{2.3F}$  29.4,  $J_{2.2F}$  179.6), 98.6 (dd, C3,  $J_{3.2F}$ 28.6,  $J_{3,3F}$  177.2), 128.4, 129.4, 130.1 (ArCH), 139.2 (ArC);  $\delta_F$  (CD<sub>3</sub>OD, 376 MHz) −187.7 (ddddd, 3F, J<sub>3F,1</sub> 3.4, J<sub>3F,2F</sub> 6.9, J<sub>3F,2</sub> 13.7, J<sub>3F,4</sub> 28.6, J<sub>3F,3</sub> 50.1), −188.1 (ddddd, 2F,  $J_{2F,3F}$  6.9,  $J_{2F,3}$  17.2,  $J_{2F,1'}$  20.6,  $J_{2F,1}$  36.6,  $J_{2F,2}$ 50.4);  $m/z$  (ESI +ve) 228 ([M + H]<sup>+</sup>, 100).

2R,3R-Difluoro-2,4-imino-1,2,3,4-tetradeoxy-D-arabinofuranose, 16. Palladium on charcoal 47 (10% wt., 5 mg) was added to a solution of N-benzyl difluoro alcohol (15 mg, 0.066 mmol) in 1,4 dioxane/ $H_2O$  (1:1, 2 mL). The reaction was flushed with argon and hydrogen gas sequentially and stirred vigorously for 2 h at rt under hydrogen until mass spectrometry showed the completion of reaction  $(m/z$  (ESI +ve) 138 [ $\overline{M}$  + H]<sup>+</sup>). After filtration, the solvent was removed in vacuo to afford the title compound 16 as a clear oil (7 mg, 82%): HRMS  $m/z$  (ESI +ve) found 138.0725 [M + H]<sup>+</sup>, C<sub>5</sub>H<sub>10</sub>F<sub>2</sub>NO<sup>+</sup> requires 138.0725;  $[\alpha]_D^{20}$  +4.3 (c 0.37, MeOH);  $\nu_{\text{max}}$  (thin film) 3321 (br, m, OH);  $\delta_{\rm H}$  (D<sub>2</sub>O, 400 MHz) free base: 3.18 (1H, dddd, H1, J<sub>1,3F</sub> 2.2, J<sub>1,2</sub> 3.9, Jgem 13.9, J<sub>1,2F</sub> 36.7), 3.24 (1H, dd, H1', Jgem 13.9, J<sub>1',2F</sub> 23.3), 3.33  $(1H, ddt, H4, J_{4,3} 3.5, J_{4,5} = J_{4,5'} 6.5, J_{4,3} 27.5), 3.69 (1H, dd, H5, J_{5,4} 6.5,$  $J_{\text{gem}}$  11.6), 3.73 (1H, ddt, H5',  $J_{5',3} = J_{5',F}$  1.0,  $J_{5',4}$  6.0,  $J_{\text{gem}}$  11.7), 5.04 (1H, dddt, H3,  $J_{3,2} = J_{3,5'} 1.0$ ,  $J_{3,4} 3.5$ ,  $J_{3,2F} 16.4$ ,  $J_{3,3F} 50.1$ ),  $5.27$  (1H, dddt, H2,  $J_{2,1'} = J_{2,3}$  1.0,  $J_{2,1}$  4.1,  $J_{2,3F}$  12.5,  $J_{2,2F}$  50.0);  $\delta_C$  (D<sub>2</sub>O, 100 MHz) 50.5 (dd, C1,  $J_{1,3F}$  2.4,  $J_{1,2F}$  23.4), 61.3 (d, C5,  $J_{5,F}$  7.6), 64.9 (d, C4,  $J_{4,3F}$  23.8), 96.8 (dd, C2,  $J_{2,3F}$  30.5,  $J_{2,2F}$  174.5), 97.8 (dd, C3,  $J_{3,2F}$  29.5,  $J_{3,3F}$  177.3);  $\delta_F$ (D2O, 376 MHz) −188.5 (ddddd, 3F, J3F,1 2.3, J3F,2F 6.9, J3F,2 12.6, J3F,4 27.5,  $J_{3F,3}$  50.3),  $-188.9$  (ddddd, 2F,  $J_{3F,2F}$  6.9,  $J_{2F,3}$  16.0,  $J_{2F,1'}$  22.7,  $J_{2F,1}$ 35.2,  $J_{2F,2}$  50.4);  $m/z$  (ESI +ve) 138 ([M + H]<sup>+</sup>, 100).

N-Benzyl-3R,4R-difluoro-L-proline, 48. Sodium hydroxide (1 M, aq, 0.24 mL, 0.24 mmol) was added to a solution of difluoromethyl ester **28** (28 mg, 0.11 mmol) in 1,4-dioxane/ $H_2O$  (1:1, 2 mL). After the solution was stirred at 40 °C for 4 h, mass spectrometry showed the formation of product and disappearance of starting of material. Then solvent was removed in vacuo to afford a residue which was loaded onto a Serdolit CG 400 resin column (the resin was prestirred in sodium hydroxide  $(1 M, aq)$  for 15 min, and then flushed with  $H<sub>2</sub>O$  until the eluent was neutral). After washing with 1,4-dioxane and  $H_2O$ , acetic acid (2 M, aq) was used to release the product. The solvent was removed in vacuo to give the pure product 48 as a light yellow glass (17 mg, 64%): HRMS  $m/z$  (ESI +ve) found 264.0805 [M + Na]<sup>+</sup>, C<sub>12</sub>H<sub>13</sub>F<sub>2</sub>NNaO<sub>2</sub><sup>+</sup> requires 264.0807;  $[\alpha]_{\rm D}^{\;\rm 20}$  – 11.7 (c 0.16, MeOH),  $\nu_{\rm max}$  (thin film) 3030  $(OH, br)$ , 1677  $(C=O, s)$ ;  $\delta_H (D_2O, 400 \text{ MHz})$  3.96 (1H, ddt, H5, J<sub>5.4</sub> =  $J_{5,3F}$  3.1,  $J_{gem}$  14.0,  $J_{5,4F}$  40.0), 4.21 (1H, dd, H5',  $J_{gem}$  13.9,  $J_{5',4F}$  17.3), 4.56 (1H, d, H6,  $J_{gem}$  12.9), 4.70 (1H, d, H6′,  $J_{gem}$  12.9), 4.73 (1H, d, H2,  $J_{2,\mathrm{F}}$ 26.6), 5.48−5.63 (2H, m, H3, H4), 7.49−7.59 (5H, m, ArH);  $\delta_{\rm C}$  (D<sub>2</sub>O, 100 MHz) 58.3 (d, C5, J<sub>5,4F</sub> 21.0), 60.7 (C6), 72.0 (d, C2, J<sub>2,3F</sub> 23.8), 91.8 (dd, C3/C4, J 33.4, J 186.0), 95.0 (dd, C3/C4, J 31.5, J 179.3), 129.4, 129.7 (ArCH), 131.1 (ArC), 131.5 (ArCH), 167.5 (d, C1, J<sub>1.F</sub> 9.5);  $\delta_F$  (D<sub>2</sub>O, 376 MHz) −182.0 (ddddd, 3F, J<sub>3F,5</sub> 2.9, J<sub>3F,4F</sub> 6.3, J<sub>3F,4</sub> 14.3,  $J_{3F,2}$  26.9,  $J_{3F,3}$  48.2), −193.2 (ddddd, 4F,  $J_{4F,3F}$  8.0,  $J_{4F,3}$  14.9,  $J_{4F,5'}$ 17.2,  $J_{4F,5}$  39.7,  $J_{4F,4}$  47.4);  $m/z$  (ESI +ve) 242 ([ $\overline{M}$  + H]<sup>+</sup>, 100).

3R,4R-Difluoro-L-proline, 12. Palladium on charcoal (10% wt, 5 mg) was added to a solution of N-benzyldifluoroproline 48 (17 mg, 0.07 mmol) in 1,4-dioxane/ $H<sub>2</sub>O$  (1:1, 2 mL). The reaction was flushed with argon and hydrogen gas sequentially and stirred vigorously for 4 h at rt under hydrogen until mass spectrometry showed the completion of reaction  $(m/z$  (ESI –ve) 150 [M – H]<sup>-</sup>). After filtration, the solvent was removed in vacuo to afford the title compound 12 as a light yellow glass (10 mg, 86%): HRMS  $m/z$  (ESI –ve) found 150.0367 [M – H]<sup>−</sup>,  $C_5H_6F_2NO_2^-$  requires 150.0372;  $[a]_D^2$ <sup>0</sup> –4.6 (c 0.19, MeOH);  $\nu_{\text{max}}$ (thin film) 3023 (br, s, OH), 1636 (s, C=O);  $\delta_H$  (D<sub>2</sub>O, 400 MHz) 3.82  $(1H, ddt, H5, J<sub>S.4</sub> = J 3.1, J<sub>gem</sub> 14.5, J<sub>S.4F</sub> 39.1), 3.96 (1H, dd, H5', J<sub>gem</sub> 14.2,$ J5′,4F 21.8), 4.61 (1H, d, H2, J2,F 24.9), 5.42−5.54 (1H, m, H4), 5.58 (1H, dd, H3,  $J_{3,4}$  6.3,  $J_{4,F}$  47.9);  $\delta_C$  (D<sub>2</sub>O, 100 MHz) 50.3 (d, C5,  $J_{5,4F}$  22.9), 66.5 (d, C2,  $J_{2,3F}$  20.0), 92.2 (dd, C4,  $J_{4,3F}$  32.4,  $J_{4,4F}$  177.4), 95.5 (dd, C3,  $J_{3,\rm 4F}$  32.9,  $J_{3,\rm 3F}$  182.6), 168.7 (d, C1,  $J_{1,\rm F}$  12.4);  $\delta_{\rm F}$  (D<sub>2</sub>O, 376 MHz, <sup>1</sup>Hdecoupled) −185.8 (d, J 15.6), −190.5 (d, J 15.6); m/z (ESI −ve) 150  $([M - H]^{-}, 100)$ .

1,2,4,6-Tetra-O-acetyl-3-deoxy-3-fluoro-D-glucopyranoside, 29. Method 1. A solution of 21 (6.88 g, 26.24 mmol) in 1:1 water/ trifluoroacetic acid (20.5 mL) was stirred at rt for 18 h after which TLC analysis (ethyl acetate) showed complete conversion of the starting material ( $R_f$  0.97) to a single major product ( $R_f$  0.06). The mixture was concentrated in vacuo, and the crude 3-deoxy-3-fluoroglucose was used without any further purification.

Acetic acid anhydride (95.9 mL, 931.5 mmol) was added to a solution of the crude glucopyranose in pyridine (95.9 mL), and the reaction mixture was stirred at rt. After 4 h, TLC analysis (cyclohexane/ethyl acetate, 1:1) showed complete conversion of the starting material (baseline) to two major products  $(R_f 0.67$  and  $0.57)$  and a minor product  $(R_f 0.16)$ . The mixture was concentrated in vacuo, and the resulting oil was partitioned between ethyl acetate  $(130 \text{ mL})$  and  $1:1 \text{ NaHCO}_3$  (satd aq)/brine (130 mL). The aqueous layer was extracted with ethyl acetate  $(2 \times 130 \text{ mL})$ . The combined organic layers were dried (MgSO<sub>4</sub>), filtered, and concentrated in vacuo, and the residue was purified by flash chromatography (cyclohexane/ethyl acetate, 77:23) to give the title compound 29 as a white solid (8.53 g, 93%).

Method 2. A solution of  $21$  (15.28 g, 58.3 mmol) in a mixture of 1,4dioxane (75 mL) and water 30 mL was stirred with Dowex (50W X8, H+ ) at 65 °C for 18 h after which TLC analysis (ethyl acetate) showed complete conversion of the starting material  $(R_f 0.93)$  to a single major product  $(R_f 0.07)$ . The mixture was concentrated in vacuo, and the crude 3-deoxy-3-fluoroglucose was used without any further purification.

Acetic anhydride (75 mL, 931.5 mmol) was added to a solution of the crude glucopyranose (10.6 g) in pyridine (75 mL), and the reaction mixture was stirred at rt. After 18 h, TLC analysis (cyclohexane/ethyl acetate, 1:1) showed complete conversion of the starting material  $(R_f 0.00)$  to a single major product  $(R_f 0.89)$ . The mixture was concentrated in vacuo, and the residue was purified by flash chromatography  $(10\% \rightarrow 25\%$  ethyl acetate in cyclohexane) to give the title compound 29 as a white solid as a 1:1 mixture of anomers (19.75 g, 97%): HRMS  $m/z$  (ESI +ve) found 373.0910 [M + Na]<sup>+</sup>, C<sub>14</sub>H<sub>19</sub>FNaO<sub>9</sub><sup>+</sup> requires 373.0905; mp 86−88 °C;  $\nu_{\text{max}}$  (thin film) 1744 (s, C=O);  $\delta_{\text{H}}$  (CDCl<sub>3</sub>, 400 MHz) 2.05 (3H, s, CH<sub>3</sub>), 2.06 (3H, s, CH<sub>3</sub>), 2.07 (3H, s, CH<sub>3</sub>), 2.08  $(\times 2)$  (2 × 3H, 2s, CH<sub>3</sub>), 2.09 (3H, s, CH<sub>3</sub>), 2.10 (3H, s, CH<sub>3</sub>), 2.14 (3H, s, CH<sub>3</sub>), 3.72 (1H, dddd, H5 $\beta$ , J<sub>5,F</sub> 1.3, J<sub>5,6</sub> 2.2, J<sub>5,6′</sub> 4.5, J<sub>5,4</sub> 10.1), 3.98– 4.04 (1H, m, H5 $\alpha$ ), 4.08 (1H, dt, H6 $\alpha$ , J<sub>6,5</sub> = J<sub>6,F</sub> 2.1, J<sub>gem</sub> 12.5), 4.09 (1H, dt, H6 $\beta$ , J<sub>6,5</sub> = J<sub>6,F</sub> 1.9, J<sub>gem</sub> 12.6), 4.22 (1H, dd, H6′ $\alpha$ , J<sub>6′,5</sub> 4.3, J<sub>gem</sub> 12.6), 4.24 (1H, dd, H6′ $\beta$ , J<sub>6′,5</sub> 4.5, J<sub>gem</sub> 12.6), 4.59 (1H, dt, H3 $\beta$ , J<sub>3,2</sub> = J<sub>3,4</sub> 9.1,  $J_{3,\mathrm{F}}$  49.5), 4.80 (1H, dt, H3 $\alpha$ ,  $\bar{j_{3,2}}$  =  $J_{3,4}$  9.4,  $J_{3,\mathrm{F}}$  53.3), 5.14 (1H, ddd, H2 $\alpha$ ,  $J_{2,1}$  3.8,  $J_{2,3}$  9.7,  $J_{2,F}$  12.1), 5.15–5.18 (3H, m, H2 $\beta$ , H4 $\alpha$ , H4 $\beta$ ), 5.63 (1H, d, H1 $\beta$ ,  $J_{1,2}$  8.3), 6.31 (1H, d, H1 $\alpha$ ,  $J_{1,2}$  3.8);  $\delta_C$  (CDCl<sub>3</sub>, 100 MHz) 20.4 (CH<sub>3</sub>), 20.5 (×2) (CH<sub>3</sub>), 20.6 (CH<sub>3</sub>), 20.7 (CH<sub>3</sub>), 61.2 (C6 $\alpha$ , C6 $\beta$ ), 67.6 (d, C4 $\alpha$ /C4 $\beta$ , J<sub>4,F</sub> 19.0), 67.6 (d, C4 $\alpha$ /C4 $\beta$ , J<sub>4,F</sub> 18.3), 69.5 (d, C2 $\alpha$ ,  $J_{2,F}$  17.5), 69.6 (d, C5 $\alpha$ ,  $J_{5,F}$  8.0), 70.2 (d, C2 $\beta$ ,  $J_{2,F}$  19.1), 71.8 (d, C5 $\beta$ ,  $J_{5,F}$ 7.2), 90.4 (d, C1 $\alpha$ , J<sub>1,F</sub> 9.5), 89.5 (d, C3 $\alpha$ , J<sub>3,F</sub> 189.9), 91.1 (d, C1 $\beta$ , J<sub>1,F</sub> 11.1), 91.5 (d, C3β, J<sub>3,F</sub> 191.5), 168.5, 168.9 (×2), 169.1, 169.5, 170.5,  $170.6$  (C=O););  $\delta_F$  (CDCl<sub>3</sub>, 376 MHz) – 200.0 (ddt, J 3.4, J = J 12.6, J<sub>F.3</sub> 52.7),  $-196.0$  (dt,  $J_{F,2} = J_{F,4}$  12.6,  $J_{F,3}$  51.5;  $m/z$  (ESI +ve) 373 ([M + Na]<sup>+</sup>, 100). 20:1 ratio: major  $\beta$ -anomer data:  $\delta_{\rm H}$  (CDCl<sub>3</sub>, 400 MHz) 2.08 (3H, s, CH<sub>3</sub>), 2.09 (3H, s, CH<sub>3</sub>), 2.10 (3H, s, CH<sub>3</sub>), 2.11 (3H, s, CH<sub>3</sub>), 3.73 (1H, dddd, H5,  $J_{5,6}$  1.3, J 2.3,  $J_{5,6'}$  4.6, J 10.1), 4.11 (1H, dt,  $\rm H6, J_{6,5}$ 1.8,  $J_{\rm{gem}}$ 12.4), 4.26 (1H, dd,  $\rm H6', J_{6',5}$ 4.7,  $J_{\rm{gem}}$ 12.5), 4.59 (1H, dt, H3, J 9.1,  $J_{3,F}$  51.8), 5.20−5.28 (2H, m, H2, H4), 5.64 (1H, d, H1,  $J_{1,2}$ 8.3);  $\delta_C$  (CDCl<sub>3</sub>, 100 MHz) 20.6 (CH<sub>3</sub>), 20.6 (CH<sub>3</sub>), 20.7 (CH<sub>3</sub>), 20.8  $(CH<sub>3</sub>), 61.3 (C6), 67.6 (d, C2, J<sub>2,F</sub> 19.1), 70.2 (d, C4, J<sub>4,F</sub> 19.1), 71.8$ 

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(d, C5,  $J_{5,F}$  8.1), 91.1 (d, C1,  $J_{1,F}$  11.1), 91.6 (d, C3,  $J_{3,F}$  191.2), 169.0, 169.0, 169.1, 170.6  $(4 \times C=0)$ .

Methyl 2,4,6-Tri-O-acetyl-3-deoxy-3-fluoro-β-D-glucopyranoside, 30. HBr (33% in acetic acid, 38.7 mL, 213.7 mmol) was added dropwise to a solution of 29 (4.86 g, 13.87 mmol) in DCM (0.13 mL) at 0  $\degree$ C. The solution was stirred for 5 h at 0  $\degree$ C after which TLC analysis (cyclohexane/ethyl acetate, 1:1) showed complete conversion of the starting material  $(R_f 0.52)$  to a single major product  $(R_f 0.77)$ . The reaction mixture was diluted with DCM (90 mL) and poured onto ice−water (90 mL). The organic layer was washed with icecold NaHCO<sub>3</sub> (satd aq, 90 mL), and the aqueous layer was extracted with DCM (90 mL). The combined organic layers were dried (MgSO<sub>4</sub>), filtered, and concentrated in vacuo at 20 °C to give the crude bromide which was used without further purification.

Silver carbonate (6.50 g, 23.59 mmol) was added to a solution of crude bromide in methanol (62.3 mL), and the mixture was stirred at rt in the dark for 18 h after which TLC analysis (cyclohexane/ethyl acetate, 1:1) showed complete conversion of the starting material  $(R_f 0.77)$ to a single major product ( $R_f$  0.48). The reaction mixture was filtered through Celite and concentrated in vacuo, and the residue was purified by flash chromatography (cyclohexane/ethyl acetate, 77:23) to give the title compound 30 as a white solid (3.91 g, 87%): HRMS  $m/z$  (ESI +ve) found 345.0967 [M + Na]<sup>+</sup>, C<sub>13</sub>H<sub>19</sub>FNaO<sub>8</sub><sup>+</sup> requires 345.0956; mp 87– 90 °C;  $\left[\alpha\right]_{D}^{20}$  –26.7 (c 0.81, CHCl<sub>3</sub>);  $\nu_{\text{max}}$  (thin film) 1744 (s, C=O);  $\delta_H$  (CDCl<sub>3</sub>, 400 MHz) 2.09 (3H, s, CH<sub>3</sub>), 2.10 (3H, s, CH<sub>3</sub>), 2.12 (3H, CH<sub>3</sub>), 3.50 (3H, s, OMe), 3.60 (1H, a-ddq, H5,  $J_{5,6} = J = J 1.3$ ,  $J_{5,6'}$  4.7,  $J_{5,4}$ 10.0), 4.16 (1H, ddd, H6, J<sub>6,5</sub> 1.4, J<sub>6,F</sub> 2.5, J<sub>gem</sub> 12.3), 4.26 (1H, dd, H6<sup>'</sup>,  $J_{6',5}$  4.8,  $J_{\text{gem}}$  12.3), 4.35 (1H, dd, H1, J 0.7,  $J_{1,2}$  8.1), 4.54 (1H, dt, H3,  $J_{3,2} = J_{3,4}$  9.1,  $J_{3,F}$  52.2), 5.10 (1H, ddd, H2,  $J_{2,1}$  8.0,  $J_{2,3}$  9.2,  $J_{2,F}$  13.3), 5.21 (1H, ddd, H4,  $J_{4,5}$  9.1,  $J_{4,3}$  10.0,  $J_{4,F}$  12.5);  $\delta_C$  (CDCl<sub>3</sub>, 100 MHz) 20.6  $(CH<sub>3</sub>)$ , 20.7 (CH<sub>3</sub>), 20.8 (CH<sub>3</sub>), 57.0 (OCH<sub>3</sub>), 61.7 (C6), 68.3 (d, C4,  $J_{4,F}$  19.1), 70.9 (d, C5,  $J_{5,F}$  8.1), 71.2 (d, C2,  $J_{2,F}$  19.1), 91.7 (d, C3,  $J_{3,F}$ 190.3), 101.1 (d, C1,  $J_{1,F}$  11.1), 169.1, 170.7 ( $3 \times C = O$ );  $\delta_F$  (CDCl<sub>3</sub>, 376 MHz) −195.9 (dt,  $J_{F,2} = J_{F,4}$  12.8,  $J_{F,3}$  52.2);  $m/z$  (ESI +ve) 345  $([M + Na]^+, 100).$ 

Methyl 3-Deoxy-3-fluoro-β-D-glucopyranoside, 31. Sodium methoxide (0.65 g, 12.0 mmol) was added to a solution of 30 (6.45 g, 20.03 mmol) in methanol (71 mL) at rt. After 5 h, TLC analysis (ethyl acetate) showed complete conversion of the starting material  $(R_f 0.90)$ to a single major product ( $R_f$  0.33). The mixture was concentrated in vacuo, and the residue was purified by flash chromatography (ethyl acetate/methanol, 17:3) to give the title compound 31 as a colorless oil (3.77 g, 96%) which crystallized on standing: HRMS  $m/z$  (ESI +ve) found 219.0632 [M + Na]<sup>+</sup>, C<sub>7</sub>H<sub>13</sub>FNaO<sub>5</sub><sup>+</sup> requires 219.0639; mp 120– 124 °C;  $[\alpha]_D^{20}$  –34.7 (c 0.73, CH<sub>3</sub>OH) [lit.<sup>[50](#page-14-0)</sup> mp 129.5–130 °C;  $[\alpha]_D$ −33.5 (c 1.2, H<sub>2</sub>O)];  $\nu_{\text{max}}$  (thin film) 3361 (br, m, OH);  $\delta_{\text{H}}$  (CD<sub>3</sub>OD, 400 MHz) 3.27 (1H, a-ddq, H5,  $J_{5,6'} = J = J 1.2$ ,  $J_{5,6}$  5.4,  $J_{5,4}$  9.9), 3.39 (1H, ddd, H2, J<sub>2,1</sub> 7.8, J<sub>2,3</sub> 9.1, J<sub>2,F</sub> 14.2), 3.54 (3H, s, OCH<sub>3</sub>), 3.57 (1H, ddd, H4, J<sub>4,3</sub> 8.7, J<sub>4,5</sub> 9.9, J<sub>4,F</sub> 14.2), 3.70 (1H, dd, H6, J<sub>6,5</sub> 5.4, J<sub>gem</sub> 12.0), 3.87 (1H, ddd, H6',  $J_{6',5}$  1.3,  $J_{6',F}$  2.3,  $J_{gem}$  12.0), 4.20 (1H, dd, H1, J 0.7,  $J_{1,2}$ 7.8), 4.24 (1H, dt, H3,  $J_{3,2} = J_{3,4}$  8.8,  $J_{3,F}$  52.8);  $\delta_C$  (CD<sub>3</sub>OD, 100 MHz) 57.5 (OMe), 62.2 (C6), 69.7 (d, C4,  $J_{4,F}$  18.1), 73.6 (d, C2,  $J_{2,F}$  18.1), 76.7 (d, C5,  $J_{5,F}$  9.1), 98.5 (d, C3,  $J_{3,F}$  182.2), 104.6 (d, C1  $J_{1,F}$  12.1);  $\delta_F$  $(CDCl_3$ , 376 MHz) −196.1 (dt, J<sub>F,2</sub> = J<sub>F,4</sub> 14.3, J<sub>F,3</sub> 52.6); m/z (ESI +ve) 219 ([M + Na]<sup>+</sup>, 100).

Methyl 6-O-(tert-butyldimethysilyl)-3-deoxy-3-fluoro-β-Dglucopyranoside, 32. Imidazole (5.42 g, 79.6 mmol) and tertbutyldimethylsilyl chloride (5.20 g, 34.5 mmol) were added to a solution of 31 (5.20 g, 26.5 mmol) in anhydrous DMF (10.0 mL) at −20 °C. The reaction mixture was stirred for 2.5 h at  $-20\,^{\circ}\mathrm{C}$  after which the mixture was allowed to warm to rt. After 3 h, TLC analysis (cyclohexane/ethyl acetate, 1:1) showed complete conversion of the starting material  $(R_f 0.03)$  to a single major product  $(R_f 0.58)$ . The reaction mixture was diluted with ethyl acetate (100 mL) and washed with 1:1 water/brine  $(3 \times 60 \text{ mL})$ . The combined aqueous layers were extracted with ethyl acetate  $(3 \times 40 \text{ mL})$ . The combined organic layers were dried  $(MgSO<sub>4</sub>)$ , filtered, and concentrated in vacuo, and the residue was purified by flash chromatography (cyclohexane/ethyl acetate,  $9:1 \rightarrow 4:1 \rightarrow 13:7$ ) to give the title compound 32 as a white solid (7.42 g, 98%): HRMS  $m/z$ (ESI +ve) found 333.1504  $[M + Na]^+$ ,  $C_{13}H_{27}FNaO_5Si^+$  requires

333.1504; mp 80−84 °C;  $[\alpha]_D^2$ <sup>0</sup> −40.0 (c 1.27, CHCl<sub>3</sub>);  $\nu_{\text{max}}$  (thin film) 3267 (br, m, OH);  $\delta_{\rm H}$  (CDCl<sub>3</sub>, 400 MHz) 0.10 (6H, 2s, CH<sub>3</sub>Si), 0.90  $(9H, s, (CH<sub>3</sub>)CSi), 2.66$  (1H, br-s, 2-OH), 3.35 (1H, dddd, H5, J 1.0,  $J_{5,6'}$  4.8,  $J_{5,6}$  5.8, J 9.5), 3.39 (1H, br-s, 4-OH), 3.54 (3H, s, OMe), 3.58 (1H, ddd, H2, J2,1 7.8, J2,3 9.1, J2,F 13.6), 3.78−3.86 (1H, m, H4), 3.86 (1H, ddd, H6, J 0.8, J<sub>6,5</sub> 5.8, J<sub>gem</sub> 10.6), 3.96 (1H, ddd, H6', J 0.8, J<sub>6',5</sub> 4.8,  $J_{\text{perm}}$  10.6), 4.21 (1H, dd, H1,  $\check{J}$ 0.8,  $J_{1,2}$  7.8), 4.42 (1H, dt, H3,  $J_{3,4} = J_{3,2}$  8.8,  $\widetilde{J}_{3,F}$ , 52.8);  $\delta_C$  (CDCl<sub>3</sub>, 100 MHz) –5.5 (2 × CH<sub>3</sub>Si), 18.2 ((CH<sub>3</sub>)CSi), 25.8 ((CH<sub>3</sub>)CSi), 57.3 (OMe), 64.2 (C6), 71.3 (d, C4, J<sub>4,F</sub> 17.6), 72.3  $(d, C2, J_{2,F} 17.6), 73.1 (d, C5, J_{5,F} 8.0), 96.4 (d, C3, J_{3,F} 183.0), 102.8 (d,$ C1,  $J_{1,F}$  12.0);  $\delta_F$  (CDCl<sub>3</sub>, 376 MHz) −196.4 (dt,  $J_{F,2} = J_{F,4}$  13.7,  $J_{F,3}$ 52.7);  $m/z$  (ESI +ve) 310 ([M + Na]<sup>+</sup>, 100).

Methyl N-Benzyl-6-O-(tert-butyldimethylsilyl)-3-fluoro-2,4 imino-2,3,4-trideoxy-β-p-talopyranoside, 23. Triflic anhydride (16.1 mL, 95.7 mmol) was added to a solution of pyridine (15.5 mL, 191 mmol) and diol 32 (7.42 g, 23.9 mmol) in DCM (100 mL) at −30 °C. The reaction mixture was stirred at −30 to −10 °C for 2 h after which time TLC analysis (cyclohexane/ethyl acetate, 3:1) showed almost complete conversion of the starting material  $(R_f 0.17)$  to a single major product ( $R_f$  0.67). A further portion of pyridine (3.86 mL, 47.8 mmol) and triflic anhydride (4.0 mL, 23.9 mmol) was added, and the reaction stirred for a further 1 h at −30 °C after which the reaction mixture was diluted with DCM (80 mL) and washed with 2 M HCl  $(100 \text{ mL})$  and the aqueous layer was extracted with DCM  $(2 \times 50 \text{ mL})$ . The combined organic layers were dried  $(MgSO<sub>4</sub>)$ , filtered, and concentrated in vacuo to give the crude ditriflate 22 (13.73 g, quant). [Partial data:  $\delta_{\rm H}$  (CDCl<sub>3</sub>, 400 MHz) 0.10 (6H, 2s, CH<sub>3</sub>Si), 0.92 (9H, s, (CH3)CSi), 3.55−3.58 (1H, m, H5), 3.59 (3H, s, OMe), 3.89 (1H, dd, H6,  $J_{6,5}$  3.3,  $J_{\text{gem}}$  12.1), 3.98 (1H, dd, H6',  $J_{6',5}$  2.1,  $J_{\text{gem}}$  12.0), 4.49 (1H, d, H1,  $J_{1,2}$  8.1), 4.69 (1H, ddd, H2,  $J_{2,1}$  7.8,  $J_{2,3}$  9.1,  $J_{2,F}$  12.9), 4.83 (1H, dt, H3,  $J_{3,4} = J_{3,2}$  8.9,  $J_{3,F}$  51.6), 5.16 (1H, ddd, H4,  $J_{4,3}$  9.0,  $J_{4,5}$  9.6,  $J_{4,F}$  12.1);  $\delta_C$  (CDCl<sub>3</sub>, 100 MHz) –5.6 (CH<sub>3</sub>Si), –5.5 (CH<sub>3</sub>Si), 18.3 ((CH<sub>3</sub>)<sub>3</sub>CSi),  $25.7 ((CH<sub>3</sub>)<sub>3</sub>CSi)$ , 57.7 (OMe), 60.3 (C6), 72.7 (d, C5,  $J<sub>5F</sub>$  6.0), 78.1 (d, C4,  $J_{4,F}$  18.1), 81.8 (d, C2,  $J_{2,F}$  18.1), 89.6 (d, C3,  $J_{3,F}$  196.2), 99.8 (d, C1,  $J_{1,F}$  9.1).]

Benzylamine (13.0 mL, 120 mmol) and N,N-diisopropylethylamine (10.2 mL, 59.8 mmol) were added to a solution of the crude ditriflate (13.73 g) in acetonitrile (140 mL), and the solution was heated to 65 °C. After 18 h, TLC analysis (cyclohexane/ethyl acetate, 3:1) showed complete conversion of the starting material  $(R_f 0.72)$  to a single major product  $(R_f 0.80)$ . The reaction mixture was concentrated and the residue was purified by flash chromatography ( $1\% \rightarrow 5\% \rightarrow 10\%$ , ethyl acetate in cyclohexane) to give the title compound 23 as a light yellow oil (7.6 g, 84%): HRMS  $m/z$  (ESI +ve) found 404.2019 [M + Na]<sup>+</sup>, ,  $C_{20}H_{32}$ FNNa $O_3S_1$ <sup>+</sup> requires 404.2028;  $[\alpha]_D^{20}$  –47.1 (c 1.15, CHCl<sub>3</sub>);  $\nu_{\text{max}}$  (thin film) fingerprint region only;  $\delta_{\text{H}}$  (CDCl<sub>3</sub>, 400 MHz) −0.05  $(3H, s, CH_3Si), -0.01$   $(3H, s, CH_3Si), 0.82$   $(9H, s, (CH_3)_3CSi), 3.47$  $(3H, s, \text{OMe})$ , 3.51 (1H, ddd, H2,  $J_{2,1}$  1.3,  $J_{2,4}$  6.0,  $J_{2,F}$  13.2), 3.68 (1H, ddd, H4,  $J_{4.5}$  1.2,  $J_{4.2}$  6.0,  $J_{4. F}$  13.3), 3.78 (1H, ddd, H6,  $J_{6. F}$  1.7,  $J_{6.5}$  5.4,  $J_{gem}$ 9.5), 3.84 (1H, dd, H6',  $J_{6'$ , 5 8.2,  $J_{\text{gem}}$  9.2), 3.94 (1H, ddt, H5,  $J_{5,4} = J_{5,\text{F}}$  1.2,  $J_{5,6}$  5.4,  $J_{5,6'}$  8.2), 4.26 (2H, br-s, CH<sub>2</sub>Ph), 4.81 (1H, t, H1,  $J_{1,2} = J_{1,F}$  1.2), 5.07 (1H, d, H3, J<sub>3,F</sub> 58.4), 7.19−7.24 (1H, m, ArH), 7.27−7.32 (2H, m, ArH), 7.39–7.43 (2H, m, ArH);  $\delta_C$  (CDCl<sub>3</sub>, 100 MHz) –5.5 (2 × CH<sub>3</sub>Si), 18.2 ((CH<sub>3</sub>)CSi), 25.8 ((CH<sub>3</sub>)CSi), 56.0 (CH<sub>2</sub>Ph), 56.1  $(OMe)$ , 63.1 (C6), 65.1 (d, C4,  $J_{4,F}$  18.3), 67.7 (d, C2,  $J_{2,F}$  18.3), 77.2 (d, C5,  $J_{5,F}$  6.4), 98.3 (d, C3,  $J_{3,F}$  213.0), 100.8 (d, C1,  $J_{1,F}$  11.1), 126.8, 128.2, 128.3 (ArCH), 139.0 (ArC);  $\delta_F$  (CDCl<sub>3</sub>, 376 MHz) −197.0 (dt, J<sub>F.2</sub> =  $J_{F,4}$  13.2,  $J_{F,3}$  58.4);  $m/z$  (ESI +ve) 382 ([M + H]<sup>+</sup>, 100).

5,6-Di-O-acetyl-N-benzyl-3-fluoro-2,4-imino-2,3,4-trideoxy-D-talose Acetyl Methyl Acetal, 33. Boron trifluoride diethyl etherate  $(4.5 \text{ mL}, 36.7 \text{ mmol})$  was added dropwise to a solution of 23  $(3.11 \text{ g})$ 8.2 mmol) in acetic anhydride (30 mL) at −30 °C before warming to rt. After 18 h, mass spectrometry (ESI +ve) showed the formation of the desired product  $412 [M + H^+]$  and no remaining starting material. The mixture was concentrated under reduced pressure, and the residue was dissolved in ethyl acetate (40 mL) and washed with  $NaHCO<sub>3</sub>$  (satd aq,  $3 \times 20$  mL). The organic fraction was dried (MgSO<sub>4</sub>), filtered, and concentrated under reduced pressure. Purification by flash column chromatography (10%  $\rightarrow$  30% ethyl acetate in cyclohexane) gave the title mixed acetal 33 (3.37 g, quant) as a pale yellow oil in a 1:1 ratio of

diastereoisomers: HRMS  $m/z$  (ESI +ve) found 412.1775 [M + H]<sup>+</sup>, ,  $C_{20}H_{27}FNO<sub>7</sub><sup>+</sup>$  requires 412.1766;  $\nu_{\text{max}}$  (thin film) 1738 (s, C=O);  $\delta_{\text{H}}$  $(CDCl_3, 400 \text{ MHz})$  1.93 (3H, s, CH<sub>3</sub>), 1.94 (3H, s, CH<sub>3</sub>), 1.97 (3H, s,  $CH<sub>3</sub>$ ), 2.07 (3H, s, CH<sub>3</sub>), 2.08 (3H, s, CH<sub>3</sub>), 2.18 (3H, s, CH<sub>3</sub>), 3.36 (1H, dt, H2,  $J_{2,1} = J_{2,3}$  5.1,  $J_{2,F}$  22.5), 3.37 (1H, dt, H2,  $J_{2,1} = J_{2,3}$  4.7,  $J_{2,F}$ 22.5), 3.39 (3H, s, OCH<sub>3</sub>), 3.41–3.52 (2H, m, H4), 3.43 (3H, s, OCH<sub>3</sub>), 3.77 (1H, d, CH<sub>2</sub>Ph, J<sub>gem</sub> 13.2), 3.80 (1H, d, CH<sub>2</sub>Ph, J<sub>gem</sub> 13.0), 3.86 (1H, d, CH<sub>2</sub>Ph, J<sub>gem</sub> 13.7), 3.87 (1H, d, CH<sub>2</sub>Ph, J<sub>gem</sub> 13.2), 4.05 (1H, dd, H6,  $J_{6,5}$  6.2,  $J_{gem}$  12.1), 4.11 (1H, dd, H6,  $J_{6,5}$  6.2,  $J_{gem}$  12.1), 4.29 (1H, dd, H6′,  $J_{6',5}$  3.7,  $J_{gem}$  12.0), 4.35 (1H, dd, H6 $^{\prime}$ , J $_{6',5}$  3.8,  $J_{gem}$  12.1), 4.85 (1H, dt, H3,  $J_{3,2} = J_{3,4}$  4.8,  $J_{3,F}$  55.9), 4.89 (1H, dt, H3,  $J_{3,2} = J_{3,4}$  4.7,  $J_{3,F}$  55.7), 4.89– 4.94 (1H, m, H5), 5.00 (1H, ddd, H5,  $J_{5,6'}$  3.8,  $J_{5,6}$  6.0,  $J_{5,4}$  7.2), 5.46 (1H, d, H1,  $J_{1,2}$  4.9), 5.66 (1H, d, H1,  $J_{1,2}$  5.1), 7.25−7.39 (10H, m, ArH);  $\delta_{\rm C}$  $(CDCl_3, 100 MHz)$  20.7 (3  $\times$  CH<sub>3</sub>), 20.8 (CH<sub>3</sub>), 20.9 (CH<sub>3</sub>), 21.0  $(CH_3)$ , 57.2 (OMe), 57.7 (OMe), 60.9 (CH<sub>2</sub>Ph), 61.6 (CH<sub>2</sub>Ph), 62.2  $(2 \times C6)$ , 67.7 (d, C4,  $J_{4,F}$  20.7), 68.2 (d, C4,  $J_{4,F}$  20.7), 69.2 (d, C2,  $J_{2,F}$ 20.7), 69.9 (d, C2,  $J_{2,F}$  19.9), 71.4 (d, C5,  $J_{5,F}$  25.4), 71.5 (d, C5,  $J_{5,F}$  25.4), 83.5 (d, C3,  $J_{3,F}$  211.6), 83.5 (d, C3,  $J_{3,F}$  211.4), 97.4 (d, C1,  $J_{1,F}$  19.9), 97.5 (d, C1, J1,F 19.9), 127.6, 127.7, 128.5 (x 2), 128.9, 129.3 (ArCH), 136.6, 137.2 (ArC), 170.0, 170.1, 170.5, 170.6 (x 2), 170.7 (C=O);  $\delta_F$ (CDCl<sub>3</sub>, 376 MHz) −179.8 (dt, J<sub>F,4</sub> = J<sub>F,5</sub> 22.9, J<sub>F,3</sub> 56.1), −180.0 (dt,  $J_{F,4} = J_{F,5}$  22.9,  $J_{F,3}$  56.1);  $m/z$  (ESI +ve) 412 ([M + H]<sup>+</sup>, 100), 434  $([M + Na]<sup>+</sup>, 40).$ 

N-Benzyl-4-fluoro-3,5-imino-3,4,5-trideoxy-D-altritol, 34. DI-BALH (1.5 M in toluene, 6.1 mL, 9.2 mmol) was added dropwise to a solution of 33 (471 mg, 1.1 mmol) in DCM (5 mL) at −78 °C. After 1.5 h, mass spectrometry (ESI +ve) showed the formation of the desired product 254  $[M + H^+]$  or gemdiol 272  $[M + H^+]$  and no remaining starting material. The reaction mixture was diluted with ethyl acetate (20 mL) and stirred with sodium potassium tartrate solution (25 mL, satd aq) until two layers were formed. The layers were separated, and the aqueous layer was extracted with ethyl acetate  $(3 \times 15 \text{ mL})$ . The combined organic layers were dried (MgSO4), filtered, and concentrated in vacuo.

Sodium borohydride (48 mg, 1.3 mmol) was added to a solution of the crude in methanol (6 mL), and the reaction mixture was stirred for 30 min. Mass spectrometry showed complete conversion of the starting material and the formation of the triol. The mixture was quenched with a few drops of glacial acetic acid and subsequently concentrated in vacuo. The crude triol was purified by ion exchange chromatography on Dowex (50W X8, H+ ) washing with dioxane and water and eluting with 2 M  $NH<sub>3</sub>$  and 1:1, 2 M NH<sub>3</sub>/dioxane. The ammoniacal fractions were concentrated in vacuo to give the triol 34 as a colorless oil (284 mg, 97%).

Large Scale. DIBALH (1.5 M in toluene, 19.7 mL, 29.5 mmol) was added dropwise to a solution of 33 (1.52 g, 3.7 mmol) in DCM (16 mL) at −78 °C. After 1.5 h, mass spectrometry (ESI +ve) showed the formation of the desired product 254  $\rm [M + H^{+}]$  or gemdiol 272  $\rm [M + H^{+}]$  and no remaining starting material. The reaction mixture was diluted with ethyl acetate (30 mL) and stirred with sodium potassium tartrate solution (80 mL, satd aq) for 2 h when two layers were formed. The layers were separated, and the aqueous layer was extracted with ethyl acetate ( $3 \times 30$  mL). The combined organic layers were dried (MgSO<sub>4</sub>), filtered, and concentrated in vacuo.

Sodium borohydride (154 mg, 4.1 mmol) was added to a solution of the crude in methanol (20 mL), and the reaction mixture was stirred for 30 min after which mass spectrometry showed complete conversion of the starting material and the formation of the triol. The mixture was quenched with a few drops of glacial acetic acid and subsequently concentrated in vacuo. The crude triol was purified by ion-exchange chromatography on Dowex (50W X8, H<sup>+</sup>) washing with dioxane and water and eluting with 2 M NH<sub>3</sub> and 1:1, 2 M NH<sub>3</sub>/dioxane. The ammoniacal fractions were concentrated in vacuo to give the triol 34 as a colorless oil (806 mg, 85%): HRMS m/z (ESI +ve) found 278.1170  $[M + Na]$ <sup>+</sup>, C<sub>13</sub>H<sub>18</sub>FNNaO<sub>3</sub><sup>+</sup> requires 278.1163. Data for HCl salt:  $[\alpha]_{D}^{20}$  –12.0 (c 0.85, CH<sub>3</sub>OH);  $\nu_{\text{max}}$  (thin film) 3299 (s, OH);  $\delta_{H}$  $(CD_3OD, 400 MHz)$  3.22 (1H, dd, H6,  $J_{6,5}$  3.9,  $J_{gem}$  12.8), 3.64 (1H, dd,  $H1, \tilde{J}_{1,2}$  5.4,  $\tilde{J}_{gem}$  11.5), 3.68 (1H, dd, H1',  $\tilde{J}_{1',2}$  4.2,  $\tilde{J}_{gem}$  11.6), 3.71 (1H, dd,  $H6', J_{6',5}$  4.3,  $J_{gem}$  12.8), 4.10–4.15 (1H, m, H2), 4.50 (1H, d, CH<sub>2</sub>Ph,  $J_{gem}$ 13.0), 4.51−4.61 (2H, m, H3, H5), 4.68 (1H, d, CH2Ph, Jgem 13.0), 5.25 (1H, dt, H4,  $J_{4,3} = J_{4,5}$  5.8,  $J_{4,F}$  56.3),7.49–7.64 (5H, m, ArH);  $\delta_C$ 

 $(CD_3OD, 100 MHz)$  58.2 (d, C6,  $J_{6,F}$  2.4), 59.9 (CH<sub>2</sub>Ph), 63.5 (C1), 69.7 (d, C2, J2,F 3.2), 72.5 (d, C5, J 25.4), 74.4 (d, C3, J 25.4), 83.5 (d, C4,  $J_{4,F}$  209.0), 130.1 (ArC), 130.5, 131.5, 132.7 (ArCH);  $\delta_F$  (CD<sub>3</sub>OD, 376 MHz)  $-183.9$  (dt,  $J_{F,5} = J_{F,3}$  18.9,  $J_{F,4}$  56.1). Data for free base:  $[\alpha]_{D}^{-20}$ +36.7 (c 0.78, CH<sub>3</sub>OH);  $\nu_{\text{max}}$  (thin film) 3366 (s, OH);  $\delta_{\text{H}}$  (CD<sub>3</sub>OD, 400 MHz) 3.12−3.18 (1H, m, H6), 3.15−3.23 (1H, m, H5), 3.22−3.27  $(1H, m, H6'), 3.33 (1H, dt, H3, J_{3,2} = J_{3,4} 5.1, J_{3,F} 29.3), 3.50 (1H, dd, H1,$  $J_{1,2}$  6.5,  $J_{\text{gem}}$  11.3), 3.55 (1H, dd, H1',  $J_{1',2}$  5.4,  $J_{\text{gem}}$  11.3), 3.64 (1H, d,  $CH_2Ph, J_{gem}$  12.4), 3.66 (1H, dt, H2,  $J_{2,1'} = J_{2,3}$  5.3,  $J_{2,1}$  6.5), 4.03 (1H, d, CH<sub>2</sub>Ph,  $J_{\text{gem}}$  12.5), 4.80 (1H, dt, H4,  $J_{4,3} = J_{4,5}$  4.8,  $J_{4,F}$  57.2), 7.25–7.38 (5H, m, ArH);  $\delta_C$  (CD<sub>3</sub>OD, 100 MHz) 62.4 (d, C6, J<sub>6F</sub> 4.0), 63.3  $(CH_2Ph)$ , 64.3 (C1), 71.2 (d, C5,  $J_{5,F}$  19.1), 71.7 (d, C3,  $J_{3,F}$  19.9), 73.4  $(d, C2, J_{2,F} 4.8), 86.7 (d, C4, J_{4,F} 207.4), 128.6, 129.4, 130.8 (ArH), 138.6)$ (ArC);  $\delta_F$  (CD<sub>3</sub>OD, 376 MHz) −182.0 (dt, J<sub>F,5</sub> = J<sub>F,3</sub> 24.0, J<sub>F,4</sub> 57.2);  $m/z$  (ESI +ve) 256 ([M + H]<sup>+</sup>, 100).

4-Fluoro-3,5-imino-3,4,5-trideoxy-D-altritol, 14. Palladium on charcoal (10% wt., 5 mg) was added to a solution of 34 (23 mg, 0.091 mmol) in 1,4-dioxane/water (1:2, 1.5 mL). The reaction mixture was flushed with argon and subsequently flushed with hydrogen. The mixture was stirred for 18 h after which mass spectrometry showed the formation of the product and no remaining starting material. The reaction mixture was filtered through (GF/B glass microfiber) and concentrated in vacuo. The residue was loaded (1:2 1,4-dioxane/water) on a Dowex (50W X8, H<sup>+</sup>) column which was prewashed with water until the pH was neutral. The column was washed with water, 1,4 dioxane, 2 M ammonia, and 1:1 1,4-dioxane/2 M ammonia. The ammoniacal fractions were concentrated in vacuo to give the title compound 14 as a light yellow oil (17 mg, 88%): HRMS  $m/z$  (ESI +ve) found 188.0693  $[M + Na]$ <sup>+</sup>, C<sub>6</sub>H<sub>12</sub>FNNaO<sub>3</sub><sup>+</sup> requires 188.0693. Data for HCl salt:  $[a]_{D}^{\ 20}$  – 35.3 (c 0.86, CH<sub>3</sub>OH);  $\nu_{max}$  (thin film) 3334 (s, br, OH, NH);  $\delta_H$  (CD<sub>3</sub>OD, 400 MHz) 3.61 (1H, dd, H<sub>1</sub>, J<sub>1,2</sub> 5.4, J<sub>gem</sub> 11.2), 3.68 (1H, dd, H1′, J<sub>1′,2</sub> 4.2, J<sub>gem</sub> 11.2), 3.89 (1H, dd, H6, J<sub>6,5</sub> 4.4, J<sub>gem</sub> 12.7), 3.94 (1H, dd, H6',  $J_{6',5}$  5.1,  $J_{gem}$  12.7), 4.04 (1H, dt, H2,  $J_{2,1'}$  4.1,  $J_{2,1}$  =  $J_{2,3}$ 5.4), 4.50−4.60 (1H, m, HS), 4.64 (1H, ddd, H3, J<sub>3,2</sub> 4.3, J<sub>3,4</sub> 5.6, J<sub>3,F</sub> 18.7), 5.33 (1H, dt, H4,  $J_{4,3} = J_{4,5}$  5.9,  $J_{4,F}$  55.9);  $\delta_C$  (CD<sub>3</sub>OD, 100 MHz) 58.7 (d, C6, J<sub>6,F</sub> 3.2), 63.9 (C1), 66.3 (d, C5, J<sub>5,F</sub> 25.5), 67.1 (d, C3, J<sub>3,F</sub> 25.5), 68.9 (d, C2,  $J_{2,F}$  3.2), 85.6 (d, C4,  $J_{4,F}$  209.0);  $\delta_F$  (CD<sub>3</sub>OD, 376 MHz) −183.5 (dt, J<sub>F,5</sub> = J<sub>F,3</sub> 18.3, J<sub>F,4</sub> 56.1); m/z (ESI +ve) 166  $([M + H]^+, 100).$ 

Methyl N-Benzyl-3-fluoro-2,4-imino-2,3,4-trideoxy-Dribonate, 24D. Sodium periodate (928 mg, 4.3 mmol) was added to a solution of 34 (922 mg, 3.6 mmol) in aqueous acetone (2:1, water/ acetone, 36 mL) and was stirred at rt for 1 h. TLC analysis (ethyl acetate) showed complete conversion of the starting material  $(R_f 0.56)$ to a single major product  $(R_f 0.74)$ . Ethanol (40 mL) was added to the reaction mixture and stirred for 2 h after which the resultant precipitate was removed via filtration (GF/B glass microfibre). The filtrate was concentrated in vacuo at 20 °C to give the crude aldehyde (803 mg).

Potassium carbonate (1.5 g, 10.8 mmol) was added to a solution of the crude aldehyde (803 mg) in methanol (40 mL). Iodine (1.19 g, 4.7 mmol) was dissolved (sonication) in methanol (40 mL) and was subsequently added dropwise to the reaction mixture at 0 °C. The reaction mixture was stirred for 1 h at 0 °C. TLC analysis (cyclohexane/ ethyl acetate, 1:1) showed complete conversion of the starting material  $(R_f 0.02)$  to a single major product  $(R_f 0.56)$ . The reaction was quenched with  $\text{Na}_2\text{SO}_3$  (satd aq) until the solution was colorless, the mixture was partitioned between ethyl acetate (100 mL) and water (100 mL), and the aqueous fraction was extracted with ethyl acetate  $(3 \times 60 \text{ mL})$ . The combined organic layers were dried (MgSO4), filtered, and concentrated in vacuo, and the residue was purified by flash chromatography ( $10\% \rightarrow$ 67%, ethyl acetate in cyclohexane) to give the title compound 24D as a brown oil (627 mg, 69%): HRMS m/z (ESI +ve) found 276.1003  $[M + Na]<sup>+</sup>$ , C<sub>13</sub>H<sub>16</sub>FNNaO<sub>3</sub><sup>+</sup> requires 276.1006;  $[\alpha]_D^{20}$  +50.0 (c 0.5, CHCl<sub>3</sub>);  $\nu_{\text{max}}$  (thin film) 3450 (w, br, OH), 1740 (s, C=O);  $\delta_{\text{H}}$  $(CDCl<sub>3</sub>$ , 400 MHz) 2.44 (1H, br-s, OH), 3.17 (1H, dd, H5, J<sub>5.4</sub> 2.8, J<sub>gem</sub> 12.1), 3.37 (1H, ddddd, H4, J 0.6, J<sub>4,5</sub>' 2.1, J<sub>4,5</sub> 2.8, J<sub>4,3</sub> 4.9, J<sub>4,F</sub> 21.5), 3.43 (1H, dd, H5',  $J_{5/4}$  1.7,  $J_{\text{gem}}$  12.2), 3.70 (3H, s, OCH<sub>3</sub>), 3.73 (1H, d, CH<sub>2</sub>Ph  $J_{\text{gem}}$  12.5), 3.78 (1H, dd, H2,  $J_{2,3}$  4.9,  $J_{2,F}$  22.3), 3.98 (1H, d, CH<sub>2</sub>Ph,  $\tilde{J}_{\text{gem}}$  12.5), 5.05 (1H, dt, H3,  $J_{3,4} = J_{3,5}$  4.9,  $J_{3,F}$  56.0), 7.26–7.36 (5H, m, ArH);  $\delta_C$  (CDCl<sub>3</sub>, 100 MHz) 52.1 (OCH<sub>3</sub>), 60.1 (d, C5, <span id="page-13-0"></span> $J_{5,F}$  4.0), 60.5 (CH<sub>2</sub>Ph), 67.9 (d, C2,  $J_{2,F}$  21.5), 69.7 (d, C4,  $J_{4,F}$  20.7), 84.4 (d, C3, J3,F 214.6), 128.0, 128.6, 129.3 (ArCH), 135.9 (ArC), 170.2 (d, C1,  $J_{1,F}$  4.8);  $\delta_F$  (CDCl<sub>3</sub>, 376 MHz) −181.5 (dt,  $J_{F,2} = J_{F,4}$  22.3,  $J_{F,3}$ 55.7);  $m/z$  (ESI +ve) 254 ([M + H]<sup>+</sup>, 100), 276 ([M + Na]<sup>+</sup>, 50).

N-Benzyl-3-fluoro-2,4-imino-2,3,4-trideoxy-D-ribonic Acid, 35D. Potassium carbonate (16 mg, 0.114 mmol) was added to a solution of 24D (22 mg, 0.087 mmol) in 1:2 1,4-dioxane/water (1.6 mL). The solution was heated to 40 °C and stirred for 18 h after which TLC analysis (cyclohexane/ethyl acetate, 1:1) showed complete conversion of the starting material  $(R_f 0.71)$  to a single major product  $(R_f 0.0)$ . HCl (2 M, 0.2 mL) was added to the reaction mixture until pH 4, and the solvent was removed in vacuo. The residue was loaded (1:2, 1,4 dioxane/water) onto a Dowex (50W X8, H<sup>+</sup>) column, which was prewashed with water, 1,4-dioxane and water sequentially. The column was eluted with water followed by 1,4-dioxane after which the product was released with ammonia (2 M). The ammoniacal fraction was concentrated in vacuo to give the title compound 35D as a light yellow glass (12 mg, 57%): HRMS  $m/z$  (ESI +ve) found 262.0853 [M + Na]<sup>+</sup>, ,  $C_{12}H_{14}$ FNNa $O_3^+$  requires 262.0850;  $[\alpha]_D^{20}$  +14.1 (c 0.4, H<sub>2</sub>O);  $\nu_{\text{max}}$ (thin film) 3227 (w, br, OH), 1631 (s, C=O);  $\delta_{\rm H}$  (Py- $d_{\rm S}$ , 400 MHz) 3.62 (1H, dq, H4,  $J_{4,3} = J_{4,5} = J_{4,5'}$  4.6,  $J_{4,F}$  22.3), 3.73 (1H, dd, H5,  $J_{5,4}$  4.9,  $J_{\text{gem}}$  11.7), 3.76 (1H, dd, H5′,  $J_{S'/4}$  4.3,  $J_{\text{gem}}$  11.5), 3.95 (1H, d, CH<sub>2</sub>Ph,  $J_{\text{gem}}$ 13.0), 4.15 (1H, dd, H2,  $J_{2,3}$  5.0,  $J_{2,F}$  23.4), 4.36 (1H, d, CH<sub>2</sub>Ph,  $J_{\text{gem}}$ 13.0), 5.62 (1H, dt, H3,  $J_{3,2} = J_{3,4}$  5.0,  $J_{3,\text{F}}$  57.1), 6.76 (br-s, OH), 7.21– 7.60 (5H, m, ArH);  $\delta_C$  (Py- $d_5$ , 126 MHz) 61.4 (CH<sub>2</sub>Ph), 62.5 (C5), 69.4 (d, C2,  $J_{2,F}$  20.0), 70.7 (d, C4,  $J_{4,F}$  19.1), 88.0 (d, C3,  $J_{3,F}$  211.7), 128.2, 129.0, 130.6 (ArCH), 137.8 (ArC), 173.8 (d, C1,  $J_{1,F}$  3.8);  $\delta_F$  (Py-d<sub>5</sub>, 376 MHz) −178.5 (dt,  $J_{F,2} = J_{F,4}$  22.9,  $J_{F,3}$  57.2);  $m/z$  (ESI +ve) 240 ([M + H]<sup>+</sup>, 100), 262 ([M + Na]<sup>+</sup>, 50); *m/z* (ESI −ve) 238 ([M − H]<sup>+</sup> , 100).

3-Fluoro-2,4-imino-2,3,4-trideoxy-D-ribonic Acid, 10D. Palladium on charcoal (10% wt., 5 mg) was added to a solution of 35D (24 mg, 0.010 mmol) in 1,4-dioxane/water (1:2, 2 mL). The reaction mixture was flushed with argon and subsequently flushed with hydrogen. The mixture was stirred for 18 h after which mass spectrometry showed the reaction went to completion. The reaction mixture was filtered through (GF/A glass microfiber) and concentrated in vacuo. The residue was loaded  $(1{:}2, 1,\!4{\text{-}}\text{dioxane}/\text{water})$  on a  $\text{Dowex}\left(\text{50W X8},\text{H}^+\right)$ column which was prewashed with water until the pH was neutral. The column was washed with water, 1,4-dioxane, 2 M ammonia, and 1:1 1,4 dioxane/2 M ammonia. The ammoniacal fractions were concentrated in vacuo to give the title compound 10D as a light yellow glass (6.6 mg, 45%): HRMS  $m/z$  (ESI –ve) found 148.0414 [M – H]<sup>-</sup>, C<sub>5</sub>H<sub>7</sub>FNO<sub>3</sub><sup>-</sup> requires 148.0415;  $[\alpha]_{D}^{\text{20}}$  +23.3 (c 0.27, H<sub>2</sub>O);  $\nu_{\text{max}}$  (thin film) 3236 (w, br, OH), 1631 (s, C=O);  $\delta_{\rm H}$  (D<sub>2</sub>O, 400 MHz) 3.91 (1H, dd, H5,  $J_{5,4}$  3.8,  $J_{gem}$  13.2), 3.98 (1H, dd, H5',  $J_{5',4}$  4.1,  $J_{gem}$  13.2), 4.67 (1H, dq, H4,  $J_{4,3} = J_{4,5} = J_{4,5'}$  4.2,  $J_{4,F}$  19.2), 4.93 (1H, dd, H2,  $J_{2,3}$  4.9,  $J_{2,F}$  21.3), 5.32 (1H, dt, H3,  $J_{3,2} = J_{3,4}$  4.7,  $J_{3,F}$  56.1);  $\delta_C$  (D<sub>2</sub>O,126 MHz) 57.9 (d, C5,  $J_{5,F}$ 3.8), 63.8 (d, C2,  $J_{2,F}$  23.8), 65.0 (d, C4,  $J_{4,F}$  26.7), 87.9 (d, C3,  $J_{3,F}$  210.8), 170.3 (d, C1,  $J_{1,F}$  4.8);  $\delta_F$  (D<sub>2</sub>O, 470 MHz) −178.3 (ddd,  $J_{F,4}$  19.3,  $J_{F,2}$ 21.3,  $J_{F,3}$  56.0);  $m/z$  (ESI –ve) 148 ([M – H]<sup>-</sup>, 100).

### **ASSOCIATED CONTENT**

# **6** Supporting Information

Copies of NMR spectra ( ${}^{1}H, {}^{13}C$  and  ${}^{19}F)$  and inhibition tables. This material is available free of charge via the Internet at [http://](http://pubs.acs.org/) [pubs.acs.org/.](http://pubs.acs.org/)

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# Notes

The authors declare no competing financial interest.

## ■ ACKNOWLEDGMENTS

This work was supported in part by the Leverhulme Trust (G.W.J.F.) and a Grant-in-Aid for Scientific Research (C) (No. 26460143) (A.K.) from the Japanese Society for the Promotion of Science (JSPS).

#### ■ REFERENCES

(1) (a) Wang, J.; Sanchez-Rosello, M.; Acena, J. L.; del Pozo, C.; Sorochinsky, A. E.; Fustero, S.; Soloshonok, V. A.; Liu, H. Chem. Rev. 2014, 114, 2432–2506. (b) Tysoe, C.; Withers, S. G. Curr. Topic Med. Chem. 2014, 14, 865−874. (c) Marsh, E. N. G.; Suzuki, Y. ACS Chem. Biol. 2014, 9, 1242−1250. (d) Buer, B. C.; Marsh, E. N. G. Protein Sci. 2012, 21, 453−462. (e) Hagmann, W. K. J. Med. Chem. 2008, 51, 4359− 4369. (f) Liang, T.; Neumann, C. N.; Ritter, T. Angew. Chem., Int. Ed. 2013, 52, 8214−8264. (g) Card, P. J. J. Carbohydr. Chem. 1984, 4, 451− 487. (h) Champagne, P.; Desroches, J.; Paquin, J.-F. Synthesis 2015, 47, 306−322.

(2) (a) Kern, N.; Felten, A.-S.; Weibel, J.-M.; Pale, P.; Blanc, A. Org. Lett. 2014, 16, 6104−6107. (b) Reddy, B. V. S.; Kishore, C.; Reddy, A. S. Tetrahedron Lett. 2014, 55, 49−51. (c) Couty, F.; Evano, G. Synlett 2009, 3053−3064. (d) Jasinski, M.; Moreno-Clavijo, E.; Reissig, H.-U. Eur. J. Org. Chem. 2014, 442−454. (e) Stankovic, S.; D'Hooghe, M.; Vanderhaegen, T.; Tehrani, K. A.; De Kimpe, N. Synlett 2014, 75−80. (f) Lo, C.; David, O.; Couty, F. Tetrahedron Lett. 2014, 55, 535−537. (g) Hodgson, D. M.; Mortimer, C. L.; McKenna, J. M. Org. Lett. 2015, 17, 330−333.

(3) (a) Risseeuw, M. D. P.; Overhand, M.; Fleet, G. W. J.; Simone, M. I. Amino Acids 2013, 45, 613−689. (b) Drouillat, B.; Peggion, C.; Wright, K.; Couty, F.; Formaggio, F.; Toniolo, C. J. Pept. Sci. 2014, 20, S189− S190. (c) Zukauskaite, A.; Moretto, A.; Peggion, C.; De Zotti, M.; Sackus, A.; Formaggio, F.; De Kimpe, N.; Mangelinckx, S. Eur. J. Org. Chem. 2014, 2312−2321. (d) Sun, Y.; Cao, Z.; Gou, S.; Hu, T. T. Chem. Biodiversity 2014, 11, 115−125. (e) Lensen, N.; Marais, J.; Brigaud, T. Org. Lett. 2015, 17, 342−345.

(4) (a) Pizzonero, M.; Dupont, S.; Babel, M. J. Med. Chem. 2014, 57, 10044−10057. (b) Rzasa, R. M.; Frohn, M. J.; Andrews, K. L. Bioorg. Med. Chem. 2014, 22, 6570−6585. (c) Phillips, D. P.; Gao, W.; Yang, Y. J. Med. Chem. 2014, 57, 3263−3282. (d) Pan, S.; Gray, N. S.; Gao, W. ACS Med. Chem. Lett. 2013, 4, 333−337. (e) Parsy, C.; Alexandre, F.-R.; Brandt, G. Bioorg. Med. Chem. Lett. 2014, 24, 4444−4449. (f) Metkar, S. D.; Bhatia, M. S.; Desai, U. V. Med. Chem. Res. 2013, 22, 5982−5989. (g) Rajulu, G. G.; Naik, H. S. B.; Kumar, G. C. Med. Chem. Res. 2014, 23, 2856−2868. (h) Alam, M. P.; Khdour, O. M.; Arce, P. M. Bioorg. Med. Chem. 2014, 22, 4935−4947. (i) Takhi, M.; Sreenivas, K.; Reddy, C. K. Eur. J. Med. Chem. 2014, 84, 382−394. (j) Hickey, E. R.; Zindell, R.; Cirillo, P. F.; Wu, L. F.; Ermann, M.; Berry, A. K.; Thomson, D. S.; Albrecht, C.; Gemkowc, M. J.; Riether, D. Bioorg. Med. Chem. Lett. 2015, 25, 575−580.

(5) Search on Feb 17, 2015, on [Web of Science](www.webofknowledge.com) for patents with azetidine in the title from Jan 1, 2013.

(6) Fowden, L. Nature 1955, 176, 347−348.

(7) (a) Rubenstein, E. J. Neuropathol. Exp. Neurol. 2008, 67, 1035− 1040. (b) Vranova, V.; Rejsek, K.; Formanek, P. Listy Cukrov. Reparske 2012, 128, 22−25.

(8) Baek, S. H.; Lee, J. G.; Park, S. Y.; Piao, X. L.; Kim, H. Y.; Bae, O. N.; Park, J. H. J. Food Compos. Anal. 2012, 25, 137−141.

(9) Bessonov, K.; Bamm, V. V.; Harauz, G. Phytochemistry 2010, 71, 502−507.

(10) (a) Rubenstein, E. Medicine 2000, 79, 80−89. (b) Dasuri, K.; Ebenezer, P. J.; Uranga, R. M.; Gavilan, E.; Zhang, L.; Fernandez-Kim, S. O. K.; Bruce-Keller, A. J.; Keller, J. N. J. Neurosci. Res. 2011, 89, 1471− 1477.

(11) Rubenstein, E.; McLaughlin, T.; Winant, R. C.; Sanchez, A.; Eckart, M.; Krasinska, K. M.; Chien, A. Phytochemistry 2009, 70, 100− 104.

(12) (a) Rodgers, K. J. Exptl. Neorology 2014, 253, 192−196. (b) Rubenstein, E. Azetidine-2-Carboxylic Acid and Other Nonprotein Amino Acids in the Pathogenesis of Neurodevelopmental Disorders in

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Comprehensive Developmental Neuroscience. In Neural Circuit Development And Function In The Healthy And Diseased Brain; Rubenstein, J. L. R., Rakic P., Eds.; Academic Press: New York, 2013; pp 539−545.

(13) (a) Shibasaki, T.; Sakurai, W.; Hasegawa, A.; Uosaki, Y.; Mori, H.; Yoshida, M.; Ozaki, A. Tetrahedron Lett. 1999, 40, 5227−5230. (b) Chang, D.; Feiten, H.-J.; Engesser, K.-H.; van Beilen, J. B.; Witholt, B.; Li, Z. Org. Lett. 2002, 4, 1859−1862.

(14) Glawar, A. F. G.; Jenkinson, S. F.; Thompson, L.; Nakagawa, S.; Kato, A.; Butters, T. D.; Fleet, G. W. J. ChemMedChem. 2013, 8, 658− 666.

(15) (a) Buku, A.; Faulstich, H.; Wieland, T.; Dabrowski, J. Proc. Natl. Acad. Sci. U.S.A. 1980, 77, 2370−2371. (b) Nakajima, T.; Volcani, B. E. Science 1969, 164, 1400−1401.

(16) Loenarz, C.; Sekirnik, R.; Thalhammer, A.; Ge, W.; Spivakovsky, E.; Mackeen, M. M.; McDonough, M. A.; Cockman, M. E.; Kessler, B. M.; Ratcliffe, P. J.; Wolf, A.; Schofield, C. J. Proc. Natl. Acad. Sci. U.S.A. 2014, 111, 4019−4024.

(17) Ayers, B. J.; Glawar, A. F. G.; Martínez, R. F.; Ngo, N.; Liu, Z.; Fleet, G. W. J.; Butters, T. D.; Nash, R. J.; Yu, C.-Y.; Wormald, M. R.; Nakagawa, S.; Adachi, I.; Kato, A.; Jenkinson, S. F. J. Org. Chem. 2014, 79, 3398−3409.

(18) Weids, A. J.; Grant, C. M. J. Cell Sci. 2014, 127, 1327−1335. (b) Hara, R.; Uchiumi, N.; Okamoto, N.; Kino, K. Biosci. Biotechnol. Biochem. 2014, 78, 1384−1388.

(19) Kollonit, J.; Barash, L.; Doldoura, G. A. J. Am. Chem. Soc. 1970, 92, 7494−7495.

(20) Van Hende, E.; Verniest, G.; Deroose, F.; Thuring, J.-W.; Macdonald, G.; Kimpe, N. J. Org. Chem. 2009, 75, 2250−2253.

(21) (a) Powell, N. H.; Clarkson, G. J.; Notman, R.; Raubo, P.; Martin, N. G.; Shipman, M. Chem. Commun. 2014, 50, 8797−8800. (b) Kiss, L.; Fueloep, F. Chem. Rev. 2014, 114, 1116−1169. (c) Zhao, J.; Wang, J. J. Phys. Chem. B 2015, 119, 3387−3397. (d) Martinek, T. A.; Fueloep, F. Chem. Rev. 2012, 41, 687−702. (e) Claridge, T. D. W.; Goodman, J. M.; Moreno, A.; Angus, D.; Barker, S. F.; Taillefumier, C.; Watterson, M. P.; Fleet, G. W. J. Tetrahedron Lett. 2001, 42, 4251−4255.

(22) (a) Rjabovs, V.; Turks, M. Tetrahedron 2013, 69, 10693−10710. (b) Chakraborty, T. K.; Kumar, N. V. S.; Roy, S.; Dutta, S. K.; Kunwar, A. C.; Sridhar, B.; Singh, H. J. Phys. Org. Chem. 2011, 24, 720−731. (c) Chakraborty, T. K.; Srinivasu, P.; Tapadar, S.; Mohan, B. K. J. Chem. Sci. 2004, 116, 187−207. (d) Chakraborty, T. K.; Arora, A.; Roy, S.; Kumar, N.; Maiti, S. J. Med. Chem. 2007, 50, 5539−5542. (e) Smith, M. D.; Claridge, T. D. W.; Tranter, G. E.; Sansom, M. S. P.; Fleet, G. W. J. J. Chem. Soc., Chem. Commun. 1998, 2041−2042.

(23) (a) Asano, N. Glycobiology 2003, 13, 93R−104R. (b) Asano, N. Cell. Molec. Life Sci. 2009, 66, 1479−1492. (b) Nash, R. J.; Kato, A.; Yu, C.-Y.; Fleet, G. W. J. Future Med. Chem. 2011, 3, 1513−1521.

(24) Lenagh-Snow, G. M. J.; Araujo, N.; Jenkinson, S. F.; Rutherford, C.; Nakagawa, S.; Kato, A.; Yu, C.-Y.; Weymouth-Wilson, A. C.; Fleet, G. W. J. Org. Lett. 2011, 13, 5834−5837.

(25) Lenagh-Snow, G. M. J.; Araujo, N.; Jenkinson, S. F.; Martinez, R. F.; Shimada, Y.; Yu, C.-Y.; Kato, A.; Fleet, G. W. J. Org. Lett. 2012, 14, 2142−2145.

(26) Martínez, R. F.; Fleet, G. W. J. Tetrahedron: Asymmetry 2014, 25, 373−380.

(27) Backer, D. C.; Horton, D.; Tindal, C. G. Carbohydr. Res. 1972, 24, 192−197.

(28) Kim, D. W.; Ahn, D.-S.; Oh, Y.-H.; Lee, S.; Kil, H. S.; Oh, S. J.; Lee, S. J.; Kim, J. S.; Ryu, J. S.; Moon, D. H.; Chi, D. Y. J. Am. Chem. Soc. 2006, 128, 16394−16397.

(29) Egli, M.; Pallan, P. S.; Allerson, C. R.; Prakash, T. P.; Berdeja, A.; Yu, J. H.; Lee, S.; Watt, A.; Gaus, H.; Bhat, B.; Swayze, E. E.; Seth, P. P. J. Am. Chem. Soc. 2010, 133, 16642−16649.

(30) Manta, S.; Tzioumaki, N.; Tsoukala, E.; Panagiotopoulou, A.; Pelecanou, M.; Balzarini, J.; Komiotis, D. Eur. J. Med. Chem. 2009, 44, 4764−4771.

(31) Kováč, P.; Yeh, H. J. C.; Glaudemans, C. P. J. Carbohydr. Res. 1987, 169, 23−34.

(32) Best, D.; Chairatana, P.; Glawar, A. F. G.; Crabtree, E.; Butters, T. D.; Wilson, F. X.; Yu, C.-Y.; Wang, W.-B.; Jia, Y.-M.; Adachi, I.; Kato, A.; Fleet, G. W. J. Tetrahedron Lett. 2010, 51, 2222−2224.

(33) Cohen, S.; Levy, D.; Bergmann, E. D. Chem. Ind. 1964, 43, 1802− 1803.

(34) (a) Mahe, O.; L'Heureux, A.; Couturier, M.; Bennett, C.; Clayton, S.; Tovell, D.; Beaulieu, F.; Paquin, J. F. J. Fluorine Chem. 2013, 153, 57− 60.

(35) Anxionnat, B.; Robert, B.; George, P.; Ricci, G.; Perrin, M. A.; Pardo, D. G.; Cossy, J. J. Org. Chem. 2012, 77, 6087−6099.

(36) D'Hooghe, M.; Catak, S.; Stankovic, S.; Waroquier, M.; Kim, Y.; ̧ Ha, H.-J.; Van Speybroeck, V.; De Kimpe, N. Eur. J. Org. Chem. 2010, 4920−4931.

(37) For details of assays, see: Glawar, A. F. G.; Best, D.; Ayers, B.; Miyauchi, S.; Nakagawa, S.; Aguilar-Moncayo, M.; García Fernández, J. M.; Mellet, C. O.; Crabtree, E. V.; Butters, T. D.; Wilson, F. X.; Kato, A.; Fleet, G. W. J. Chem.—Eur. J. 2012, 18, 9341−9359 and references cited therein.

(38) In the [Supporting Information,](#page-13-0) Table 1 compares the fluoroazetidines with their oxygenated analogues and Table 2 shows the effect of the N-benzyl derivatives in Figure [2](#page-4-0).

(39) Kato, A.; Kato, N.; Miyauchi, S.; Minoshima, Y.; Adachi, I.; Ikeda, K.; Asano, N.; Watson, A. A.; Nash, R. J. Phytochemistry 2008, 69, 1261-1265.

(40) Lee, J. C.; Francis, S.; Dutta, D.; Gupta, V.; Yang, Y.; Zhu, J.-Y.; Tash, J. S.; Schonbrunn, E.; Georg, G. I. J. Org. Chem. 2012, 77, 3082− 3098.

(41) (a) Andersen, S. M.; Ebner, M.; Ekhart, C. W.; Gradnig, G.; Legler, G.; Lundt, I.; Stütz, A. E.; Withers, S. G.; Wrodnigg, T. Carbohydr. Res. 1997, 301, 155−166. (b) Albert, R.; Dax, K.; Seidl, S.; Sterk, H.; Stütz, A. E. J. Carbohydr. Chem. 1985, 4, 513−520. (c) Prell, E.; Csuk, R. Bioorg. Med. Chem. Lett. 2009, 19, 5673−5674.

(42) Li, Y.I-X.; Shimada, Y.; Sato, K.; Kato, A.; Zhang, W.; Jia, Y.-M.; Fleet, G. W. J.; Xiao, M.; Yu, C.-Y. Org. Lett. 2015, 17, 716−729.

(43) PANC-1 cells were purchased from the Japanese Collection of Research Bioresources (JCRB). Cells were cultured in DMEM supplemented with 1× MEM nonessential amino acid medium (Sigma), antibiotic antimycotic solution (Sigma), 4.5 g/L-glucose, 110 mg/mL sodium pyruvate, and 10% fetal calf serum (MP Biomedicals). (44) (a) Carmichael, J.; DeGraff, W.; Gazdar, A. F.; Minna, J. D.; Mitchell, J. B. Cancer Res. 1987, 47, 943−946. (b) Awale, S.; Lu, J.; Kalauni, S. K.; Kurashima, Y.; Tezuka, Y.; Kadota, S.; Esumi, H. Cancer Res. 2006, 66, 1751−1757. (c) Kaji, A.; Saito, R.; Nomura, M.; Miyamoto, K.; Kiriyama, N. Anticancer Res. 1997, 17, 3675−3679.

(45) Gottlieb, H. E.; Koplyar, V.; Nudelman, A. J. Org. Chem. 1997, 62, 7512−7515.

(46) Buck, K. W.; Foster, A. B.; Hems, R.; Webber, J. M. Carbohydr. Res. 1966, 3, 137−138.

(47) Li, Y.-X.; Huang, M.-H.; Yamashita, Y.; Kato, A.; Jia, Y.-M.; Wang, W.-B.; Fleet, G. W. J.; Nash, R. J.; Yu, C.-Y. Org. Biomol. Chem. 2011, 9, 3405−3414.

(48) Cohen, S.; Levy, D.; Bergmann, E. D. Chem. Ind. 1964, 43, 1802− 1803.

(49) Mastihubová, M.; Biely, P. Carbohydr. Res. 2004, 339, 2101− 2110.

(50) Kovac, P.; Yeh, H. J. C; Glaudemans, C. P. J. Carbohydr. Res. 1987, 169, 23−34.